Package 'POINT'

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Type Package

Title Protein Structure Guided Local Test

Version 1.2

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Description Provides an implementation of a rare variant association test that utilizes protein tertiary structure to increase signal and to identify likely causal variants. Performs structure-guided collapsing, which leads to local tests that borrow information from neighboring variants on a protein and that provide association information on a variant-specific level. For details of the implemented method see West, R. M., Lu, W., Rotroff, D. M., Kuenemann, M., Chang, S-M., Wagner M. J., Buse, J. B., Motsinger-Reif, A., Fourches, D., and Tzeng, J-Y. (2019) <doi:10.1371/journal.pcbi.1006722>.

License GPL-2

Depends methods, stats, rARPACK, Matrix, CompQuadForm

NeedsCompilation no

Encoding UTF-8

RoxygenNote 6.1.1

Collate 'A_Kernel.R' 'A_BurdenKernel.R' 'A_LinearKernel.R'
'A_PolyKernel.R' 'B_PvMethod.R' 'B_PvMethod_Davies.R'
'B_PvMethod_Liu.R' 'C_BinomialTrait.R' 'C_GaussianTrait.R'
'D_NullResult.R' 'calcLocalKernel.R' 'distanceMatrix.R'
'mainCode.R' 'point.R' 'pvResamp.R'

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Description

A rare variant association test that utilizes protein tertiary structure to increase signal and to identify likely causal variants. Performs structure-guided collapsing, which leads to local tests that borrow information from neighboring variants on a protein and that provide association information on a variant-specific level.

Details

Package:	POINT
Type:	Package
Version:	1.1
Date:	2019-03-02
License:	GPL-2
Depends:	methods, stats, rARPACK, Matrix
Imports:	CompQuadForm

Author(s)

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References

Marceau West R, Lu W, Rotroff DM, Kuenemann MA, Chang SM, et al. (2019) Identifying individual risk rare variants using protein structure guided local tests (POINT). PLOS Computational Biology 15(2): e1006722.

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point   *Protein Structure Guided Local Test*

Description

A rare variant association test that utilizes protein tertiary structure to increase signal and to identify likely causal variants. Performs structure-guided collapsing, which leads to local tests that borrow information from neighboring variants on a protein and that provide association information on a variant-specific level.

Usage

```
point(yy, X, snp, proteinCoord, ..., trait = "binomial", cValues = c(0,
  0.1, 0.2, 0.3, 0.4, 0.5), weighted = TRUE, weight = NULL,
  kernel = "linear", d = NULL, pvMethod = "davies",
  nperturb = 1000, verbose = TRUE)
```

Arguments

yy	numeric vector; phenotype values.
X	numeric matrix; non-genetic covariates.
snp	numeric matrix; genotype snp matrix (count of minor alleles). Matrix cannot contain missing values.
proteinCoord	numeric matrix; columns correspond to 3 dimensional coordinates (x,y,z) of each variant in the protein tertiary structure.
...	optional additional arguments for p-value methods CompQuadForm::davies and CompQuadForm::liu.
trait	character; type of phenotype data. Must be one of {'gaussian','binomial'} quantitative or case control data, respectively.
cValues	numeric vector; c values from which to choose the optimal neighborhood size for borrowing significant information.
weighted	logical; whether or not to weight the local kernel test using (non-distance based) weights.
weight	numeric vector (optional) If NULL and weighted is TRUE (1.0-MAF)^24. Ignored if weighted is FALSE.
kernel	character; type of local kernel to use; Must be one of {'burden', 'linear', 'polynomial'}.
d	numeric; If kernel = 'poly', d is the order of the polynomial kernel.
pvMethod	character; method of calculating the p-value of each single marker test for fixed c values. Must be one of {'davies', 'liu'}.
nperturb	numeric, number of perturbations/resamples (perturbed test statistics) to calculate p-value of minP statistic.
verbose	logical; generate progress screen prints.

Value

Returns a matrix the rows of which correspond to individual markers. Columns correspond to:
(1) minP statistic;
(2) local kernel test p-value;
(3) optimal scale value from input cValues;
(4) minor allele frequency; and
(5) single variant score test p-value.

Examples

```
# number of subjects
nsubj <- 1000

# number of markers
nm <- 5

# generate coordinates for proteins
protein <- cbind( stats::rnorm(n = nm, mean = 17.6, sd =  6.6),
                  stats::rnorm(n = nm, mean =  1.6, sd = 13.6),
                  stats::rnorm(n = nm, mean = 22.9, sd = 10.4) )

# generate snp matrix
snp <- matrix(data = rbinom(n = nsubj*nm, size = 1, p = 0.02),
              nrow = nsubj, ncol = nm)
colnames(snp) = paste0("m",1:nm)

# generate binmoial response
MAF <- colMeans(x = snp)/2
causal <- numeric(nm)
causal[c(2,4)] <- 1.0
betaG <- 0.4*abs(log10(x = MAF))*causal

#no non-genetic covariates
X <- NULL
mu <- -0.05 + snp \%*\% betaG

pryy <- exp(mu)/(1+exp(mu))
yy <- sapply(X = pryy, FUN = stats::rbinom, n = 1, size = 1)

res <- point(yy = yy, X = X, snp = snp, proteinCoord = protein,
             trait = 'binomial', cValues = c(0.1,0.2),
             weighted = TRUE, weight = NULL, kernel = 'linear',
             pvMethod = 'liu', nperturb = 100,
             verbose = FALSE)
```

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