

# Package ‘SpaCCr’

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**Type** Package

**Title** Spatial Convex Clustering

**Version** 0.1.0

**Author** John Nagorski

**Maintainer** John Nagorski<jn13@rice.edu>

**Description** Genomic Region Detection via Spatial Convex Clustering. See <<https://arxiv.org/abs/1611.04696>> for details.

**Depends** R (>= 2.10)

**License** GPL-3

**LazyData** TRUE

**LinkingTo** Rcpp, RcppArmadillo

**Imports** abind, dplyr, ggplot2, parallel, Rcpp, tidy

**RoxygenNote** 5.0.1

**Suggests** testthat

**NeedsCompilation** yes

**Repository** CRAN

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CNVPlotSeriesMeans	<i>Plot subjects' copy number data with cluster means overlaid for a single chromosome</i>
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---

### Description

Plot subjects' copy number data with cluster means overlaid for a single chromosome

### Usage

```
CNVPlotSeriesMeans(Location, X, Cluster, NSubj = 3, lowery = -1,
  upperry = 1)
```

### Arguments

Location	A vector of length p with chromosomal locations
X	A variable (p) by subject (n) data matrix
Cluster	A vector of length p with cluster labels
NSubj	A positive integer; number of randomly selected subjects to plot.
lowery, upperry	Scalars; limits for y-axis

### Examples

```
NULL
```

---

GetClusters	<i>Compute Clusters from fusions</i>
-------------	--------------------------------------

---

### Description

Compute Clusters from fusions

### Usage

```
GetClusters(V)
```

### Arguments

V	An n by p-1 data matrix
---	-------------------------

**Examples**

NULL

---

GetGammaCV	<i>Get optimal cross validated gamma values by various rules</i>
------------	--

---

**Description**

Get optimal cross validated gamma values by various rules

**Usage**

```
GetGammaCV(ErrMat, rule = 1, gamma.seq)
```

**Arguments**

ErrMat	Matrix of cross validated errors outputted by GetCVerMat
rule	A number indicating how optimal gamma should be chosen. 1 for minimum cv error, 2 for 1 standard error rule
gamma.seq	sequence of regularization parameters used for cross validation.

**Value**

A scalar. Optimal gamma selected by CV rule.

**Examples**

NULL

---

GetParBlocks	<i>Function to compute blocks for parallization; should not be called directly</i>
--------------	--

---

**Description**

Function to compute blocks for parallization; should not be called directly

**Usage**

```
GetParBlocks(X, w)
```

**Arguments**

X	An n by p data matrix
w	A vector of positive scalars of length p-1

**Examples**

```
NULL
```

---

```
hello           Hello, World!
```

---

**Description**

Prints 'Hello, world!'.

**Usage**

```
hello()
```

**Examples**

```
hello()
```

---

```
methy           A subset of methylation data for 50 subjects
```

---

**Description**

A subset of methylation data for 50 subjects

**Usage**

```
methy
```

**Format**

A data frame with 1000 rows and 53 columns. Rows are variables and columns are:

**ProbeID** Varibale ID string

**Chromosome** Chromosome Number

**Genomic\_Coordinate** Probe location on chromosome in basepairs

**TCGA-A7-A0CE-11A-21D-A10Q-05** Subject ID

**TCGA-A7-A0CH-11A-32D-A10Q-05** Subject ID

**TCGA-A7-A0DB-11A-33D-A093-05** Subject ID

**TCGA-A7-A0DC-11A-41D-A10Q-05** Subject ID

**TCGA-BH-A0AY-11A-23D-A10Q-05** Subject ID

**TCGA-BH-A0BV-11A-31D-A10Q-05** Subject ID

**TCGA-BH-A0DZ-11A-22D-A10Q-05** Subject ID

TCGA-A2-A1FV-01A-11D-A13K-05 Subject ID  
TCGA-A2-A1FW-01A-11D-A13K-05 Subject ID  
TCGA-A2-A1FX-01A-11D-A13K-05 Subject ID  
TCGA-A2-A1G0-01A-11D-A13K-05 Subject ID  
TCGA-A2-A1G1-01A-21D-A13K-05 Subject ID  
TCGA-A2-A1G4-01A-11D-A13K-05 Subject ID  
TCGA-A2-A1G6-01A-11D-A13K-05 Subject ID  
TCGA-A7-A13G-01A-11D-A13K-05 Subject ID  
TCGA-A7-A13G-01B-04D-A22R-05 Subject ID  
TCGA-A7-A13G-11A-51D-A13T-05 Subject ID  
TCGA-AO-A1KO-01A-31D-A13K-05 Subject ID  
TCGA-AO-A1KP-01A-11D-A13K-05 Subject ID  
TCGA-AO-A1KQ-01A-11D-A13K-05 Subject ID  
TCGA-AO-A1KS-01A-11D-A13K-05 Subject ID  
TCGA-AO-A1KT-01A-11D-A13K-05 Subject ID  
TCGA-AQ-A1H2-01A-11D-A13K-05 Subject ID  
TCGA-AQ-A1H3-01A-31D-A13K-05 Subject ID  
TCGA-B6-A1KC-01A-11D-A13K-05 Subject ID  
TCGA-B6-A1KC-01B-11D-A161-05 Subject ID  
TCGA-B6-A1KF-01A-11D-A13K-05 Subject ID  
TCGA-B6-A1KN-01A-11D-A13K-05 Subject ID  
TCGA-BH-A1EN-01A-11D-A13K-05 Subject ID  
TCGA-BH-A1EN-11A-23D-A13T-05 Subject ID  
TCGA-BH-A1EX-01A-11D-A13K-05 Subject ID  
TCGA-BH-A1EY-01A-11D-A13K-05 Subject ID  
TCGA-BH-A1EY-11B-21D-A13T-05 Subject ID  
TCGA-BH-A1F2-01A-31D-A13K-05 Subject ID  
TCGA-BH-A1F2-11A-32D-A13T-05 Subject ID  
TCGA-BH-A1F5-01A-12D-A13K-05 Subject ID  
TCGA-BH-A1F5-11A-43D-A13T-05 Subject ID  
TCGA-BH-A1F6-01A-11D-A13K-05 Subject ID  
TCGA-BH-A1F6-11B-94D-A13T-05 Subject ID  
TCGA-BH-A1F8-01A-11D-A13K-05 Subject ID  
TCGA-BH-A1F8-11B-21D-A13T-05 Subject ID  
TCGA-BH-A1FB-01A-11D-A13K-05 Subject ID  
TCGA-BH-A1FB-11A-33D-A13T-05 Subject ID  
TCGA-BH-A1FC-01A-11D-A13K-05 Subject ID

TCGA-BH-A1FC-11A-32D-A13T-05 Subject ID  
 TCGA-BH-A1FD-01A-11D-A13K-05 Subject ID  
 TCGA-BH-A1FD-11B-21D-A13T-05 Subject ID  
 TCGA-BH-A1FE-01A-11D-A13K-05 Subject ID  
 TCGA-BH-A1FE-06A-11D-A212-05 Subject ID  
 TCGA-BH-A1FE-11B-14D-A13T-05 Subject ID

### Source

<http://cancergenome.nih.gov/>

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MethyRegionPlot	<i>Plots methylation data by Genomic Coordinates for a given chromosomal region with cluster means overlayed for each subject.</i>
-----------------	--

---

### Description

Plots methylation data by Genomic Coordinates for a given chromosomal region with cluster means overlayed for each subject.

### Usage

```
MethyRegionPlot(X, Coord, Cluster, SubjInd = 1:3, Start, End)
```

### Arguments

X	A Subject by Probe data matrix for a single chromosome of CNV data
Coord	A vector of Genomic Coordinates for a single chromosome
Cluster	Cluster labels for each probe
SubjInd	A vector of numeric indices corresponding to the Subjects to be plotted.
Start	Genomic Coordinate minimum
End	Genomic Coordinate maximum

### Examples

```
library(dplyr)
library(tidyr)
data("methy")
methy <- methy[1:20,1:10]
Coordinates <- methy$Genomic_Coordinate
methy %>%
  tbl_df() %>%
  select(-Chromosome, -Genomic_Coordinate) %>%
  gather(Subject, Value, -ProbeID) %>%
  spread(ProbeID, Value) -> X
```

```

SubjectLabels <- X$Subject
X <- X[,-1] %>% as.matrix()
nsubj <- nrow(X)
nprobes <- ncol(X)
nweights <- choose(nprobes,2)
diff.vals <- diff(Coordinates)
too.far <- diff.vals > 20000
sig = 1/5e3
w.values <- exp(-sig*diff.vals)
w.values[too.far] = 0

verbose=TRUE
tol.base = 1e-4
tol.miss = 1e-4
max.iter.base=5000
max.iter.miss=500
ngam = 20
gamma.seq <- exp(seq(log(1e-1),log(1e1),length.out=ngam))
CVRes <- SpaCC_CV(X=t(scale(t(X),center=TRUE,scale=FALSE)),
                 w=w.values,
                 gamma.seq=gamma.seq,
                 nfolds=5,
                 nu=1/nsubj,
                 verbose=TRUE,
                 tol.base=tol.base,
                 tol.miss=tol.miss,
                 max.iter.base=max.iter.base,
                 max.iter.miss=max.iter.miss,
                 parallel=FALSE,frac = .1)
PlotCV(CVRes$ErrMat,gamma.seq = CVRes$gamma.seq,rule = 1)
best.gam <- GetGammaCV(CVRes$ErrMat,rule = 1,gamma.seq = CVRes$gamma.seq)
bo <-t(scale(t(X),center=TRUE,scale=FALSE))
bo[is.na(bo)] <- mean(bo,na.rm=TRUE)
Sol <- SpaCC_Missing(t(scale(t(X),center=TRUE,scale=FALSE)),
                   w.values,
                   gamma = best.gam,
                   nu=1/nsubj,
                   verbose=TRUE,
                   tol.base=tol.base,
                   tol.miss=tol.miss,
                   max.iter.base=max.iter.base,
                   max.iter.miss=max.iter.miss,
                   bo,
                   t(diff(t(bo))),
                   t(diff(t(bo))))

VThreshed <- Sol$V
clustsThreshed <- GetClusters(VThreshed)
NEstRegion <- length(unique(clustsThreshed$cluster))
NEstRegion
VThreshed <- ThreshV(Sol$V,X,mult = 1)
clustsThreshed <- GetClusters(VThreshed)
NEstRegion <- length(unique(clustsThreshed$cluster))
NEstRegion

```

```

start.coord <- 2e5
end.coord <- 4e5
MethyRegionPlot(X,Coordinates,clustsThreshed$cluster,SubjInd = 1:3,Start=start.coord,End=end.coord)

```

---

PlotCV

*A function for plotting cross validation errors*


---

### Description

A function for plotting cross validation errors

### Usage

```
PlotCV(ErrMat, rule = 2, gamma.seq)
```

### Arguments

ErrMat	A matrix of error outputted by SpaCC_CV
rule	An interger indicating which CV rule to choose
gamma.seq	The sequence of regularization parameters

### Examples

```

library(dplyr)
library(tidyr)
data("methy")
methy <- methy[1:20,1:10]
Coordinates <- methy$Genomic_Coordinate
methy %>%
  tbl_df() %>%
  select(-Chromosome,-Genomic_Coordinate) %>%
  gather(Subject,Value,-ProbeID) %>%
  spread(ProbeID,Value) -> X
SubjectLabels <- X$Subject
X <- X[,-1] %>% as.matrix()
nsubj <- nrow(X)
nprobes <- ncol(X)
nweights <- choose(nprobes,2)
diff.vals <- diff(Coordinates)
too.far <- diff.vals > 20000
sig = 1/5e3
w.values <- exp(-sig*diff.vals)
w.values[too.far] = 0

verbose=TRUE
tol.base = 1e-4
tol.miss = 1e-4
max.iter.base=5000
max.iter.miss=500

```



```

ngam = 20
gamma.seq <- exp(seq(log(1e-1),log(1e1),length.out=ngam))
CVRes <- SpaCC_CV(X=t(scale(t(X),center=TRUE,scale=FALSE)),
                 w=w.values,
                 gamma.seq=gamma.seq,
                 nolds=5,
                 nu=1/nsubj,
                 verbose=TRUE,
                 tol.base=tol.base,
                 tol.miss=tol.miss,
                 max.iter.base=max.iter.base,
                 max.iter.miss=max.iter.miss,
                 parallel=FALSE,frac = 1)
PlotCV(CVRes$ErrMat,gamma.seq = CVRes$gamma.seq,rule = 1)

```

---

SpaCC	<i>Base function for computing SpaCC solution for single regularization value.</i>
-------	--

---

### Description

Base function for computing SpaCC solution for single regularization value.

### Usage

```
SpaCC(X, w, gamma, nu, verbose, tol, maxiter, Uinit, Vinit, Laminit)
```

### Arguments

X	A subject (n) by probe (p) data matrix
w	A vector weights for adjacent probes. Should have length nprobes -1
gamma	A scalar value for the regularization parameter
nu	A scalar value for the step size in AMA algorithm
verbose	Logical value whether progress should be printed
tol	A scalar value for convergence tolerance.
maxiter	Maximum number of iterations
Uinit	A matrix used for warm starts with U
Vinit	A matrix used for warm start with V
Laminit	A matrix used for warm starts with Lam

### Value

An RcppArmadillo field object. Has three components, each holds the U,V, and Lam matrix for the current regularization

SpaCC\_CV

*Perform Cross Validation to select gamma/sparsity level***Description**

Perform Cross Validation to select gamma/sparsity level

**Usage**

```
SpaCC_CV(X, w, gamma.seq, nfolds = 5, nu = 1/nrow(X), verbose = FALSE,
  tol.base = 1e-04, tol.miss = 1e-04, max.iter.base = 5000,
  max.iter.miss = 500, parallel = FALSE, frac = 1)
```

**Arguments**

X	A subject (n) by variable (p) matrix; the data
w	A vector of length p-1; weights for clustering
gamma.seq	A vector of positive scalars; regularization parameter sequence
nfolds	A positive scalar; number of cross validation folds
nu	A positive scalar; augmented Lagrangian paramter
verbose	Logical; should messages be printed?
tol.base	A small positive scalar; convergence tolerance for base SpaCC problem.
tol.miss	A small positive scalar; convergence tolerance for missing data problem.
max.iter.base	A positive integer; maximum number of iterations for base SpaCC problem
max.iter.miss	A positive integer; maximum number of iterations for missing data problem
parallel	A logical; should CV paths be done in parallel?
frac	A positive scalar between 0 and 1; fraction of hold out set to utilize

**Value**

A list with elements: ErrMat - a length(gamma.seq) by nfold matrix containing error on out of fold data; SpMat - a length(gamma.seq) by nfold matrix containing sparsity levels; gamma.seq - original gamma.seq sorted largest to smallest

**Examples**

```
library(dplyr)
library(tidyr)
data("methy")
methy <- methy[1:20,1:10]
Coordinates <- methy$Genomic_Coordinate
methy %>%
  tbl_df() %>%
  select(-Chromosome, -Genomic_Coordinate) %>%
  gather(Subject, Value, -ProbeID) %>%
```

```

    spread(ProbeID,Value) -> X
    SubjectLabels <- X$Subject
    X <- X[,-1] %>% as.matrix()
    nsubj <- nrow(X)
    nprobes <- ncol(X)
    nweights <- choose(nprobes,2)
    diff.vals <- diff(Coordinates)
    too.far <- diff.vals > 20000
    sig = 1/5e3
    w.values <- exp(-sig*diff.vals)
    w.values[too.far] = 0

    verbose=TRUE
    tol.base = 1e-4
    tol.miss = 1e-4
    max.iter.base=5000
    max.iter.miss=500
    ngam = 20
    gamma.seq <- exp(seq(log(1e-1),log(1e1),length.out=ngam))
    CVRes <- SpaCC_CV(X=t(scale(t(X),center=TRUE,scale=FALSE)),
                     w=w.values,
                     gamma.seq=gamma.seq,
                     nolds=5,
                     nu=1/nsubj,
                     verbose=TRUE,
                     tol.base=tol.base,
                     tol.miss=tol.miss,
                     max.iter.base=max.iter.base,
                     max.iter.miss=max.iter.miss,
                     parallel=FALSE,frac = 1)

```

---

SpaCC\_Methy

*Performs Spatial Convex Clustering for methylation data*


---

## Description

Performs Spatial Convex Clustering for methylation data

## Usage

```

SpaCC_Methy(X, Coordinates, gamma.seq, dist.cutoff = 20000, sig = 1/5000,
            weights = NULL, center = TRUE, scale = FALSE, nolds = 5, nu = NULL,
            tol.base = 1e-04, tol.miss = 1e-04, max.iter.base = 5000,
            max.iter.miss = 500, frac = 0.1, parallel = FALSE, gam.rule = 2,
            thresh.mult = 1, thresh.value = NULL)

```

## Arguments

X                    A subject (n) by variable (p) matrix; the data

Coordinates	a vector listing genomic coordinates
gamma.seq	a vector of regularization parameters
dist.cutoff	maximum distance at which probes should be regularized
sig	positive scalar controlling spatial weight decay
weights	a vector of spatial weights
center	should data be centered
scale	should data be scaled
nfolds	number of folds for cross validation
nu	parameter for augmented lagrangian
tol.base	tolerance level for base function
tol.miss	tolerance for missing function
max.iter.base	maximum number of iterations for base function
max.iter.miss	maximum number of iterations for missing function
frac	fraction of fold to use for cross validation
parallel	should algorithm be run in parallel
gam.rule	cross validation rule
thresh.mult	multiplier for threshold value
thresh.value	value of threshold

### Value

Labels a vector of cluster labels

### Examples

```

data("methy")
methy <- methy[1:20,1:10]
library(dplyr)
library(tidyr)
Coordinates <- methy$Genomic_Coordinate
methy %>%
  tbl_df() %>%
  select(-Chromosome, -Genomic_Coordinate) %>%
  gather(Subject, Value, -ProbeID) %>%
  spread(ProbeID, Value) -> X
SubjectLabels <- X$Subject
X <- X[,-1] %>% as.matrix()
verbose=TRUE
tol.base = 1e-4
tol.miss = 1e-4
max.iter.base=5000
max.iter.miss=500
ngam = 20
gamma.seq <- exp(seq(log(1e-1), log(1e1), length.out=ngam))
ClusterLabels <- SpaCC_Methy(X = X, Coordinates = Coordinates, gamma.seq = gamma.seq)

```

SpaCC\_Missing

*Solve Spatial Convex Clustering problem for missing data***Description**

Solve Spatial Convex Clustering problem for missing data

**Usage**

```
SpaCC_Missing(X, w, gamma, nu = 1/nrow(X), verbose = FALSE,
  tol.base = 1e-04, tol.miss = 1e-04, max.iter.base = 5000,
  max.iter.miss = 500, Uinit, Vinit, Laminit)
```

**Arguments**

X	A subject (n) by variable (p) matrix; the data
w	A vector of length p-1; weights for clustering
gamma	A positive scalar; regularization parameter
nu	A positive scalar; augmented Lagrangian parameter
verbose	Logical; should messages be printed?
tol.base	A small positive scalar; convergence tolerance for base SpaCC problem.
tol.miss	A small positive scalar; convergence tolerance for missing data problem.
max.iter.base	A positive integer; maximum number of iterations for base SpaCC problem
max.iter.miss	A positive integer; maximum number of iterations for missing data problem
Uinit	An n by p matrix; initial value for U
Vinit	An n by p-1 matrix; initial value for V
Laminit	An n by p-1 matrix; initial value for Lam

**Value**

A list with elements U, V, and Lam

**Examples**

```
library(dplyr)
library(tidyr)
data("methy")
methy <- methy[1:20,1:10]
Coordinates <- methy$Genomic_Coordinate
methy %>%
  tbl_df() %>%
  select(-Chromosome, -Genomic_Coordinate) %>%
  gather(Subject, Value, -ProbeID) %>%
  spread(ProbeID, Value) -> X
SubjectLabels <- X$Subject
```

```

X <- X[,-1] %>% as.matrix()
X[1:5,1:5]
nsubj <- nrow(X)
nprobes <- ncol(X)
nweights <- choose(nprobes,2)
diff.vals <- diff(Coordinates)
too.far <- diff.vals > 20000
sig = 1/5e3
w.values <- exp(-sig*diff.vals)
w.values[too.far] = 0

verbose=TRUE
tol.base = 1e-4
tol.miss = 1e-4
max.iter.base=5000
max.iter.miss=500
bo <-t(scale(t(X),center=TRUE,scale=FALSE))
bo[is.na(bo)] <- mean(bo,na.rm=TRUE)
best.gam = 1
Sol <- SpaCC_Missing(t(scale(t(X),center=TRUE,scale=FALSE)),
                    w.values,
                    gamma = best.gam,
                    nu=1/nsubj,
                    verbose=TRUE,
                    tol.base=tol.base,
                    tol.miss=tol.miss,
                    max.iter.base=max.iter.base,
                    max.iter.miss=max.iter.miss,
                    bo,
                    t(diff(t(bo))),
                    t(diff(t(bo))))

```

---

SpaCC\_Path

*Solve Spatial Convex Clustering problem for path of regularization parameters*


---

### Description

Solve Spatial Convex Clustering problem for path of regularization parameters

### Usage

```

SpaCC_Path(X, w, gamma.seq, nu = 1/nrow(X), verbose = FALSE,
           tol.base = 1e-04, tol.miss = 1e-04, max.iter.base = 5000,
           max.iter.miss = 500)

```

### Arguments

X                    A subject (n) by variable (p) matrix; the data

w	A vector of length p-1; weights for clustering
gamma.seq	A vector of positive scalars; regularization parameter sequence
nu	A positive scalar; augmented Lagrangian parameter
verbose	Logical; should messages be printed?
tol.base	A small positive scalar; convergence tolerance for base SpaCC problem.
tol.miss	A small positive scalar; convergence tolerance for missing data problem.
max.iter.base	A positive integer; maximum number of iterations for base SpaCC problem
max.iter.miss	A positive integer; maximum number of iterations for missing data problem

**Value**

A list with elements UPath, VPath, LamPath, and gamma.seq

**Examples**

NULL

---

SpaCC_Path_Parallel	<i>Solve Spatial Convex Clustering problem for path of regularization parameters in parallel</i>
---------------------	--

---

**Description**

Solve Spatial Convex Clustering problem for path of regularization parameters in parallel

**Usage**

```
SpaCC_Path_Parallel(X, w, gamma.seq, nu = 1/nrow(X), verbose = FALSE,
  tol.base = 1e-04, tol.miss = 1e-04, max.iter.base = 5000,
  max.iter.miss = 500, ncores = 2)
```

**Arguments**

X	A subject (n) by variable (p) matrix; the data
w	A vector of length p-1; weights for clustering
gamma.seq	A vector of positive scalars; regularization parameter sequence
nu	A positive scalar; augmented Lagrangian parameter
verbose	Logical; should messages be printed?
tol.base	A small positive scalar; convergence tolerance for base SpaCC problem.
tol.miss	A small positive scalar; convergence tolerance for missing data problem.
max.iter.base	A positive integer; maximum number of iterations for base SpaCC problem
max.iter.miss	A positive integer; maximum number of iterations for missing data problem
ncores	A positive integer; number of cores to use

**Value**

A list with elements UPath, VPath, LamPath, and gamma.seq

**Examples**

NULL

---

ThreshV

*Threshold differences*

---

**Description**

Threshold differences

**Usage**

ThreshV(V, X, mult = 1, thresh.value = NULL)

**Arguments**

V	an n x p-1 matrix of differences
X	an n x p matrix
mult	scalar to multiply standard deviation
thresh.value	optional user specified threshold value.

**Value**

VThreshed an n x p-1 matrix of thresholded differences

**Examples**

NULL



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