# Package 'RRphylo'

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**Description** Functions for phylogenetic analysis (Castiglione et al., 2018 <doi:10.1111/2041-210X.12954>). The functions perform the estimation of phenotypic evolutionary rates, identification of phenotypic evolutionary rate shifts, quantification of direction and size of evolutionary change in multivariate traits, the computation of ontogenetic shape vectors and test for morphological convergence.

License GPL-2

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# $\mathsf{R}$ topics documented:

RRphylo-package	3
angle.matrix	3
conv.map	6
cutPhylo	8
DataApes	9
DataCetaceans	10
DataFelids	11
DataOrnithodirans	11
DataSimians	12
DataUng	13
distNodes	13
evo.dir	14
fix.poly	17
getGenus	19
	20
<b>8 ,</b>	21
lollipoPlot	21
makeFossil	22
makeL	23
makeL1	24
namesCompare	25
node.paths	26
•	26
	31
<del>-</del>	33
1 7	34
F	36
<u>.</u>	38
1	39
1	41
rate.map	43
±	45
	45
č	50
RRphylo	53
1 *	
	<ul><li>53</li><li>53</li></ul>
search.conv	55 50
search.shift	58
search.trend	61
setBM	65
sig2BM	67
sizedsubtree	68
StableTraitsR	69
swapONE	70
tips	72

RRphylo-package 3

Index	
	78
treedataMatch	70
tree.merger	

# **Description**

**RRphylo** provides tools for phylogenetic comparative analysis. The main functions allow estimation of phenotypic evolutionary rates, identification of shifts in rate of evolution, quantification of direction and size of evolutionary change of multivariate traits, and computation of species ontogenetic vectors. Additionally, there are functions for simulating phenotypic data, manipulating phylogenetic trees, and retrieving information from phylogenies. Finally, there are functions to plot and test rate shifts at particular nodes.

The complete list of functions can be displayed with library(help = RRphylo). Citations to individual functions are available by typing citation("RRphylo").

#### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

angle.matrix	Ontogenetic shape vectors analysis

# **Description**

This function computes and compares ontogenetic vectors among species in a tree.

# Usage

```
angle.matrix(RR,node,Y=NULL,select.axes=c("no","yes"),
  type=c("phenotypes","rates"),cova=NULL,clus=0.5)
```

# **Arguments**

RR an object produced by RRphylo.

node the number identifying the most recent common ancestor to all the species the

user wants ontogenetic vectors be computed.

Y multivariate trait values at tips.

select.axes if "yes", Y variables are individually regressed against developmental stages

and only significant variables are retained to compute ontogenetic vectors. All

variables are retained otherwise.

4 angle.matrix

type specifies weather to perform the analysis on phenotypic ("phenotypes") or rate ("rates") vectors.

cova the covariate to be indicated if its effect on rate values must be accounted for. Contrary to RRphylo, cova needs to be as long as the number of tips in the tree. As the covariate only affects rates computation, there is no covariate to provide when type = "phenotypes".

clus the proportion of clusters to be used in parallel computing. To run the single-

threaded version of angle.matrix set clus = 0.

#### **Details**

The angle.matrix function takes as objects a phylogenetic tree (retrieved directly from an RRphylo object), including the different ontogenetic stages of each species as polytomies. Names at tips must be written as species ID and stage number separated by the underscore. The RRphylo object angle.matrix is fed with is just used to extract the dichotomized version of the phylogeny. This is necessary because node numbers change randomly at dichotomizing non-binary trees. However, when performing angle.matrix with the covariate the RRphylo object must be produced without accounting for the covariate. Furthermore, as the covariate only affects the rates computation, it makes no sense to use it when computing vectors for phenotypic variables. Once angles and vectors are computed, angle.matrix performs two tests by means of standard major axis (SMA) regression. For each species pair, the "biogenetic test" verifies whether the angle between species grows during development, meaning that the two species becomes less similar to each other during growth. The "paedomorphosis test" tells whether there is heterochronic shape change in the data. Under paedomorphosis, the adult stages of one (paedomorphic) species will resemble the juvenile stages of the other (peramorphic) species. The test regresses the angles formed by the shapes at different ontogenetic stages of a species to the shape at the youngest stage of the other in the pair, against age. Then, it tests whether the two regression lines (one per species) have different slopes, and whether they have different signs. If the regression lines point to different directions, it means that one of the two species in the pair resembles, with age, the juveniles of the other, indicating paedomorphosis. Ontogenetic vectors of individual species are further computed, in reference to the MRCA of the pair, and to the first stage of each species (i.e. intraspecifically). Importantly, the size of the ontogenetic vectors of rates tell whether the two species differ in terms of developmental rate, which is crucial to understand which process is behind paedomorphosis, where it applies. While performing the analysis, the function prints messages on-screen informing about tests results. If select.axes = "yes", informs the user about which phenotypic variables are used. Secondly, it specifies whether ontogenetic vectors to MRCA, and intraspecific ontogenetic vectors significantly differ in angle or size between species pairs. Then, for each species pair, it indicates if the biogenetic law and paedomorphosis apply.

### Value

A list containing 4 objects:

 \$regression.matrix a 'list' including 'angles between species' and 'angles between species to MRCA' matrices for all possible combinations of species pairs from the two sides descending from the MRCA. For each matrix, corresponding biogenetic and paedomorphosis tests are reported. angle.matrix 5

\$angles.2.MRCA.and.vector.size a 'data.frame' including angles between the resultant vector of species and the MRCA and the size of the resultant vector computed from species to MRCA, per stage per species.

- 3. **\$ontogenetic.vectors2MRCA** a 'data.frame' including angle, size, and corresponding x and y components, of ontogenetic vectors computed between each species and the MRCA. For both angle and size, the p-value for the difference between species pairs is reported.
- 4. **\$ontogenetic.vectors.to.1st.stage** a 'list' containing:
  - \$matrices: for all possible combinations of species pairs from the two sides descending form the MRCA, the upper triangle of the matrix contains the angles between different ontogenetic stages for the first species. The same applies to the lower triangle, but for the second species.
  - \$vectors: for all possible combinations of species pairs from the two sides descending form the MRCA, angles and sizes of ontogenetic vectors computed to the first stage of each species. For both, the p-value for the difference between the species pair is reported.

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

```
## Not run:
 data("DataApes")
 DataApes$PCstage->PCstage
 DataApes$Tstage->Tstage
 DataApes$CentroidSize->CS
 cc<- 2/parallel::detectCores()</pre>
 RRphylo(tree=Tstage,y=PCstage,clus=cc)->RR
# Case 1. without accounting for the effect of a covariate
# Case 1.1 selecting shape variables that show significant relationship with age
 # on phenotypic vectors
   angle.matrix(RR,node=72,Y=PCstage,select.axes="yes",type="phenotypes",clus=cc)
 # on rates vectors
   angle.matrix(RR,node=72,Y=PCstage,select.axes="yes",type="rates",clus=cc)
# Case 1.2 using all shape variables
 # on phenotypic vectors
   angle.matrix(RR,node=72,Y=PCstage,select.axes="no",type="phenotypes",clus=cc)
 # on rates vectors
   angle.matrix(RR,node=72,Y=PCstage,select.axes="no",type="rates",clus=cc)
# Case 2. accounting for the effect of a covariate (on rates vectors only)
# Case 2.1 selecting shape variables that show significant relationship with age
  angle.matrix(RR,node=72,Y=PCstage,select.axes="yes",type="rates", cova=CS,clus=cc)
```

6 conv.map

```
# Case 2.2 using all shape variables
angle.matrix(RR,node=72,Y=PCstage,select.axes="no",type="rates",cova=CS,clus=cc)
## End(Not run)
```

conv.map

Mapping morphological convergence on 3D surfaces

# **Description**

Given vectors of RW (or PC) scores, the function selects the RW(PC) axes which best account for convergence and maps convergent areas on the corresponding 3D surfaces.

# Usage

```
conv.map(dataset,pcs,mshape,conv=NULL, exclude=NULL,out.rem=TRUE,
    show.consensus=FALSE, plot=TRUE,col="blue",names = TRUE)
```

# **Arguments**

dataset	data frame (or matrix) with the RW (or PC) scores of the group or species to be compared.
pcs	RW (or PC) vectors (eigenvectors of the covariance matrix) of all the samples.
mshape	the Consensus configuration.
conv	a named character vector indicating convergent species as (indicated as "conv" in dataset) and not convergent species (indicated as "noconv").
exclude	integer: the index number of the RW (or PC) to be excluded from the comparison.
out.rem	logical: if TRUE triangles with outlying area difference are removed.
show.consensus	logical: if TRUE, the Consensus configuration is included in the comparison.
plot	logical: if TRUE, the pairwise comparisons are be plotted. For more than 5 pairwise comparisons, the plot is not shown.
col	character: the colour for the plot.
names	logical: if TRUE, the names of the groups or species are displayed in the 3d plot.

# **Details**

conv.map automatically builds a 3D mesh on the mean shape calculated from the Relative Warp Analysis (RWA) or Principal Component Analysis (PCA) (Schlager 2017) by applying the function vcgBallPivoting (Rvcg). conv.map further gives the opportunity to exclude some RW (or PC) axes from the analysis because, for example, in most cases the first axes are mainly related to high-order morphological differences driven by phylogeny and size variations. conv.map finds and plots the strength of convergence on 3D surfaces. An output of conv.map (if the dataset contains a number equal or lower then 5 items) is an interactive plot mapping the convergence on the 3D

conv.map 7

models. In the upper triangle of the 3D multiple layouts the rows representing the reference models and the columns the target models. On the contrary, on the lower triangle the rows correspond to the target models and the columns the reference models. In the calculation of the differences of areas we supply the possibility to find and remove outliers from the vectors of areas calculated on the reference and target surfaces. We suggest considering this possibility if the mesh may contain degenerate facets.

#### Value

The function returns a list including:

- **\$angle.compare** data frame including the real angles between the given shape vectors, the angles conv computed between vectors of the selected RWs (or PCs), the angles between vectors of the non-selected RWs (or PCs), the difference conv, and its p values.
- \$selected.pcs RWs (or PCs) axes selected for convergence.
- **\$average.dist** symmetric matrix of pairwise distances between 3D surfaces.
- **\$suface1** list of coloured surfaces, if two meshes are given, it represents convergence between mesh A and B charted on mesh A.
- **\$suface2** list of coloured surfaces, if two meshes are given, it represents convergence between mesh A and B charted on mesh B.
- **\$scale** the value used to set the colour gradient, computed as the maximum of all differences between each surface and the mean shape.

#### Author(s)

Marina Melchionna, Antonio Profico, Silvia Castiglione, Carmela Serio, Gabriele Sansalone, Pasquale Raia

# References

Schlager, S. (2017). Morpho and Rvcg-Shape Analysis in R: R-Packages for geometric morphometrics, shape analysis and surface manipulations. In: Statistical shape and deformation analysis. Academic Press. Melchionna, M., Profico, A., Castiglione, S., Serio, C., Mondanaro, A., Modafferi, M., Tamagnini, D., Maiorano, L., Raia, P., Witmer, L.M., Wroe, S., & Sansalone, G. (2021). A method for mapping morphological convergence on three-dimensional digital models: the case of the mammalian sabre-tooth. Palaeontology, 64, 573–584. doi:10.1111/pala.12542

# See Also

```
search.conv vignette; relWarps; procSym
```

```
## Not run:
data(DataSimians)
DataSimians$pca->pca
## Case 1. Convergent species only
dato<-pca$PCscores[c(1,4),]</pre>
```

8 cutPhylo

cutPhylo

Cut the phylogeny at a given age or node

# **Description**

The function cuts all the branches of the phylogeny which are younger than a specific age or node (i.e. the age of the node).

# Usage

```
cutPhylo(tree,age=NULL,node=NULL,keep.lineage=TRUE)
```

# **Arguments**

a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

the age (in terms of time distance from the recent) at which the tree must be cut
the node whose age must be used as cutting limit.

keep.lineage logical specifying whether lineages with no descendant tip must be retained (see example below). Default is TRUE.

# **Details**

When an entire lineage is cut (i.e. one or more nodes along a path) and keep.lineages = TRUE, the leaves left are labeled as "l" followed by a number.

### Value

The function returns the cut phylogeny and plots it into the graphic device. The time axis keeps the root age of the original tree. Note, tip labels are ordered according to their position in the tree.

DataApes 9

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

# See Also

```
cutPhylo vignette
```

# **Examples**

```
## Not run:
library(ape)

set.seed(22)
rtree(100)->tree
3->age

cutPhylo(tree,age=age)->t1
cutPhylo(tree,age=age,keep.lineage=FALSE)->t1
cutPhylo(tree,node=151)->t2
cutPhylo(tree,node=151,keep.lineage=FALSE)->t2

## End(Not run)
```

DataApes

Example dataset

# **Description**

Geometric morphometrics shape data regarding Apes' facial skeleton and Apes phylogentic trees (*Profico et al. 2017*).

# Usage

```
data(DataApes)
```

# **Format**

A list containing:

**\$PCstage** A data frame containing 38 shape variables for Apes' facial skull at different ontogenetic stages.

**\$PCadult** A data frame containing 3 shape variables for Apes' facial skull.

**\$Tstage** Phylogenetic tree of Apes including the different ontogenetic stages of each species as polytomies.

**\$Tadult** Phylogenetic tree of Apes.

\$CentroidSize numeric vector of Centroid Size values of 'PCstage'.

10 DataCetaceans

### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

### References

Profico, A., Piras, P., Buzi, C., Di Vincenzo, F., Lattarini, F., Melchionna, M., Veneziano, A., Raia, P. & Manzi, G. (2017). The evolution of cranial base and face in Cercopithecoidea and Hominoidea: Modularity and morphological integration. *American journal of primatology*, 79: e22721.

DataCetaceans

Example dataset

# **Description**

Cetaceans' body and brain mass, and phylogenetic tree (Serio et al. 2019).

# Usage

data(DataCetaceans)

# **Format**

A list containing:

treecet Cetaceans phylogenetic tree.

masscet numeric vector of cetaceans body masses (ln g).

brainmasseet numeric vector of cetaceans brain masses (ln g).

**aceMyst** body mass (ln g) for *Mystacodon selenensis*, used as node prior at the ancestor of the Mysticeti.

#### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

#### References

Serio, C., Castiglione, S., Tesone, G., Piccolo, M., Melchionna, M., Mondanaro, A., Di Febbraro, M., & Raia, P. (2019). Macroevolution of Toothed Whales Exceptional Relative Brain Size. *Evolutionary Biology*, 46: 332-342. doi:10.1007/s11692-019-09485-7

DataFelids 11

DataFelids

Example dataset

# **Description**

Geometric morphometrics shape data regarding felids' mandible and phylogentic tree (*Piras et al.*, 2018).

# Usage

data(DataFelids)

# **Format**

A list containing:

**\$PCscoresfel** A data frame containing 83 shape variables for felids' mandible.

**\$treefel** Phylogenetic tree of felids.

#### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

# References

Piras, P., Silvestro, D., Carotenuto, F., Castiglione, S., Kotsakis, A., Maiorino, L., Melchionna, M., Mondanaro, A., Sansalone, G., Serio, C., Vero, V. A., & Raia, P. (2018). Evolution of the sabertooth mandible: A deadly ecomorphological specialization. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 496, 166-174.

DataOrnithodirans

Example dataset

# **Description**

Ornithodirans' body mass, phylogenetic tree and locomotory type (Castiglione et al 2018).

# Usage

data(DataOrnithodirans)

12 DataSimians

# **Format**

A list containing:

treedino Ornithodirans phylogenetic tree.

massdino numeric vector of ornithodirans body masses.

statedino vector of ornithodirans locomotory type.

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

#### References

Castiglione, S., Tesone, G., Piccolo, M., Melchionna, M., Mondanaro, A., Serio, C., Di Febbraro, M., & Raia, P.(2018). A new method for testing evolutionary rate variation and shifts in phenotypic evolution. *Methods in Ecology and Evolution*, 9: 974-983.doi:10.1111/2041-210X.12954

DataSimians

Example dataset

# **Description**

The output of Procrustes superimposition as performed by the function procSym on 9 similars faces and the phylogenetic tree for such species.

# Usage

data(DataSimians)

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

DataUng 13

DataUng

Example dataset

# **Description**

Geometric morphometrics shape data regarding mandible and phylogentic tree of 'Ungulatomorpha' (*Raia et al.*, 2010).

# Usage

data(DataUng)

#### **Format**

A list containing:

**\$PCscoresung** A data frame containing 205 shape variables for mandible of 'Ungulatomorpha'.

**\$treeung** Phylogenetic tree of 'Ungulatomorpha'.

**\$stateung** vector of 'Ungulatomorpha' feeding type.

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

# References

Raia, P., Carotenuto, F., Meloro, C., Piras, P., & Pushkina, D. (2010). The shape of contention: adaptation, history, and contingency in ungulate mandibles. *Evolution*, 64: 1489-1503.

distNodes

Finding distance between nodes and tips

# **Description**

The function computes the distance between pairs of nodes, pairs of tips, or between nodes and tips. The distance is meant as both patristic distance and the number of nodes intervening between the pair.

# Usage

```
distNodes(tree,node=NULL,clus=0.5)
```

14 evo.dir

# Arguments

tree	a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.
node	either a single node/tip or a pair of nodes/tips.
clus	the proportion of clusters to be used in parallel computing. To run the single-threaded version of $distNodes$ set $clus = 0$ .

#### Value

If node is specified, the function returns a data frame with distances between the focal node/tip and the other nodes/tips on the tree (or for the selected pair only). Otherwise, the function returns a matrix containing the number of nodes intervening between each pair of nodes and tips.

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

### **Examples**

```
data("DataApes")
DataApes$Tstage->Tstage

cc<- 2/parallel::detectCores()
distNodes(tree=Tstage,clus=cc)
distNodes(tree=Tstage,node=64,clus=cc)
distNodes(tree=Tstage,node="Tro_2",clus=cc)
distNodes(tree=Tstage,node=c(64,48),clus=cc)
distNodes(tree=Tstage,node=c(64,"Tro_2"),clus=cc)</pre>
```

evo.dir

Phylogenetic vector analysis of phenotypic change

# Description

This function quantifies direction, size and rate of evolutionary change of multivariate traits along node-to-tip paths and between species.

# Usage

```
evo.dir(RR,angle.dimension=c("rates","phenotypes"),
  y.type=c("original","RR"),y=NULL,pair.type=c("node","tips"),pair=NULL,
  node=NULL,random=c("yes","no"),nrep=100)
```

evo.dir 15

### **Arguments**

RR an object produced by RRphylo.

angle.dimension

specifies whether vectors of "rates" or "phenotypes" are used.

y.type must be indicated when angle.dimension = "phenotypes". If "original", it

uses the phenotypes as provided by the user, if "RR" it uses RR\$predicted.phenotypes.

y specifies the phenotypes to be provided if y.type = "original".

pair.type either "node" or "tips". Angles are computed between each possible couple of

species descending from a specified node ("node"), or between a given couple

of species ("tips").

pair species pair to be specified if pair.type = "tips". It needs to be written as in

the example below.

node node number to be specified if pair.type = "node". Notice the node num-

ber must refer to the dichotomic version of the original tree, as produced by

RRphylo.

random whether to perform randomization test ("yes"/"no").

nrep number of replications must be indicated if random = "yes". It is set at 100 by

default.

#### **Details**

The way evo.dir computes vectors depends on whether phenotypes or rates are used as variables. RRphylo rates along a path are aligned along a chain of ancestor/descendant relationships. As such, each rate vector origin coincides to the tip of its ancestor, and the resultant of the path is given by vector addition. In contrast, phenotypic vectors are computed with reference to a common origin (i.e. the consensus shape in a geometric morphometrics). In this latter case, vector subtraction (rather than addition) will define the resultant of the evolutionary direction. It is important to realize that resultants could be at any angle even if the species (the terminal vectors) have similar phenotypes, because path resultants, rather than individual phenotypes, are being contrasted. However, the function also provides the angle between individual phenotypes as 'angle between species'. To perform randomization test (random = "yes"), the evolutionary directions of the two species are collapsed together. Then, for each variable, the median is found, and random paths of the same size as the original paths are produced sampling at random from the 47.5th to the 52.5th percentile around the medians. This way, a random distribution of angles is obtained under the hypothesis that the two directions are actually parallel. The 'angle direction' represents the angle formed by the species phenotype and a vector of 1s (as long as the number of variables representing the phenotype). This way, each species phenotype is contrasted to the same vector. The 'angle direction' values could be inspected to test whether individual species phenotypes evolve towards similar directions.

### Value

Under all specs, evo.dir returns a 'list' object. The length of the list is one if pair.type = "tips". If pair.type = "node", the list is as long as the number of all possible species pairs descending from the node. Each element of the list contains:

angle.path.A angle of the resultant vector of species A to MRCA

16 evo.dir

vector.size.species.A size of the resultant vector of species A to MRCA

angle.path.B angle of the resultant vector of species B to MRCA

vector.size.species.B size of the resultant vector of species B to MRCA

angle.between.species.to.mrca angle between the species paths resultant vectors to the MRCA

angle.between.species angle between species vectors (as they are, without computing the path)

MRCA the node identifying the most recent common ancestor of A and B

**angle.direction.A** angle of the vector of species A (as it is, without computing the path) to a fixed reference vector (the same for all species)

vec.size.direction.A size of the vector of species A

**angle.direction.B** angle of the vector of species B (as it is, without computing the path) to a fixed reference vector (the same for all species)

vec.size.direction.B size of the vector of species B

If random = "yes", results also include p-values for the angles.

#### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

```
## Not run:
 data("DataApes")
 DataApes$PCstage->PCstage
 DataApes$Tstage->Tstage
 cc<- 2/parallel::detectCores()</pre>
 RRphylo(tree=Tstage, y=PCstage, clus=cc)->RR
# Case 1. Without performing randomization test
 # Case 1.1 Computing angles between rate vectors
 # for each possible couple of species descending from node 72
   evo.dir(RR,angle.dimension="rates",pair.type="node",node=72 ,
   random="no")
 # for a given couple of species
   evo.dir(RR,angle.dimension="rates",pair.type="tips",
   pair= c("Sap_1","Tro_2"),random="no")
 # Case 1.2 computing angles between phenotypic vectors provided by the user
 # for each possible couple of species descending from node 72
   evo.dir(RR, angle.dimension="phenotypes", y.type="original",
    y=PCstage,pair.type="node",node=72,random="no")
 # for a given couple of species
   evo.dir(RR, angle.dimension="phenotypes", y.type="original",
   y=PCstage,pair.type="tips",pair=c("Sap_1","Tro_2"),random="no")
 # Case 1.3 computing angles between phenotypic vectors produced by "RRphylo"
```

fix.poly 17

```
# for each possible couple of species descending from node 72
   evo.dir(RR,angle.dimension="phenotypes",y.type="RR",
   pair.type="node",node=72,random="no")
 # for a given couple of species
   evo.dir(RR,angle.dimension="phenotypes",y.type="RR"
   pair.type="tips",pair=c("Sap_1","Tro_2"),random="no")
# Case 2. Performing randomization test
 # Case 2.1 Computing angles between rate vectors
 # for each possible couple of species descending from node 72
   evo.dir(RR,angle.dimension="rates",pair.type="node",node=72 ,
    random="yes",nrep=10)
 # for a given couple of species
    evo.dir(RR,angle.dimension="rates",pair.type="tips",
   pair= c("Sap_1","Tro_2"),random="yes",nrep=10)
 # Case 2.2 computing angles between phenotypic vectors provided by the user
  # for each possible couple of species descending from node 72
   evo.dir(RR,angle.dimension="phenotypes",y.type="original",
   y=PCstage,pair.type="node",node=72,random="yes",nrep=10)
 # for a given couple of species
    evo.dir(RR,angle.dimension="phenotypes",y.type="original",
   y=PCstage,pair.type="tips",pair=c("Sap_1","Tro_2"),random="yes",nrep=10)
 # Case 2.3 computing angles between phenotypic vectors produced by "RRphylo"
 # for each possible couple of species descending from node 72
   evo.dir(RR,angle.dimension="phenotypes",y.type="RR",
   pair.type="node",node=72,random="yes",nrep=10)
 # for a given couple of species
   evo.dir(RR, angle.dimension="phenotypes", y.type="RR",
   pair.type="tips",pair=c("Sap_1","Tro_2"),random="yes",nrep=10)
## End(Not run)
```

fix.poly

Resolving polytomies to non-zero length branches

# **Description**

The function either collapses clades under a polytomy or resolves polytomous clades to non-zero length branches, dichotomous clades.

# Usage

```
fix.poly(tree,type=c("collapse","resolve"),node=NULL,tol=1e-10,random=TRUE)
```

18 fix.poly

#### **Arguments**

tree a phylogenetic tree.

type either 'collapse' to create polytomies to one or more specific nodes or 'resolve'

to resolve (fix) all the polytomies within the tree or to one or more specific

nodes.

node the node in the tree where a polytomy should be created or fixed, either. If

type='resolve' and node=NULL all the polytomies present in the tree are re-

solved.

tol the tolerance to consider a branch length significantly greater than zero, set at

1e-10 by default. If type='resolve', all the branch lengths smaller than tol

are treated as polytomies.

random a logical value specifying whether to resolve the polytomies randomly (the de-

fault) or in the order they appear in the tree (if random = FALSE).

# **Details**

Under type='resolve' polytomous clades are resolved adding non-zero length branches to each new node. The evolutionary time attached to the new nodes is partitioned equally below the dichotomized clade.

### Value

A phylogenetic tree with randomly fixed (i.e. type='resolve') polytomies or created polytomies (i.e. type='collapse').Note, tip labels are ordered according to their position in the tree.

# Author(s)

Silvia Castiglione, Pasquale Raia, Carmela Serio

### References

Castiglione, S., Serio, C., Piccolo, M., Mondanaro, A., Melchionna, M., Di Febbraro, M., Sansalone, G., Wroe, S., & Raia, P. (2020). The influence of domestication, insularity and sociality on the tempo and mode of brain size evolution in mammals. *Biological Journal of the Linnean Society*,132: 221-231. doi:10.1093/biolinnean/blaa186

#### See Also

```
fix.poly vignette;
```

```
## Not run:
    require(ape)

data("DataCetaceans")
DataCetaceans$treecet->treecet

# Resolve all the polytomies within Cetaceans phylogeny
```

getGenus 19

```
fix.poly(treecet,type="resolve")->treecet.fixed
par(mfrow=c(1,2))
plot(treecet,no.margin=TRUE,show.tip.label=FALSE)
plot(treecet.fixed,no.margin=TRUE,show.tip.label=FALSE)

# Resolve the polytomies pertaining the genus Kentriodon
fix.poly(treecet,type="resolve",node=221)->treecet.fixed2
par(mfrow=c(1,2))
plot(treecet,no.margin=TRUE,show.tip.label=FALSE)
plot(treecet.fixed2,no.margin=TRUE,show.tip.label=FALSE)

# Collapse Delphinidae into a polytomous clade
fix.poly(treecet,type="collapse",node=179)->treecet.collapsed
par(mfrow=c(1,2))
plot(treecet,no.margin=TRUE,show.tip.label=FALSE)
plot(treecet.collapsed,no.margin=TRUE,show.tip.label=FALSE)
## End(Not run)
```

getGenus

Taxonomic inspection of the tree at the genus level

# **Description**

The function returns the most recent common ancestor and the number of species belonging to each or some user-specified genera within the phylogenetic tree.

# Usage

```
getGenus(tree,genera=NULL)
```

# **Arguments**

tree a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

Generic name and specific epithet must be separated by '\_'.

genera a character vector including one or more genera to focus on. Please notice the

function is case sensitive.

# Value

The function returns a data-frame including the number of species and the most recent common ancestor of each genera.

#### Author(s)

Silvia Castiglione, Pasquale Raia, Carmela Serio

20 getMommy

# **Examples**

```
DataCetaceans$treecet->tree
getGenus(tree)
getGenus(tree,c("Mesoplodon","Balaenoptera"))
```

getMommy

Upward tip or node to root path

# **Description**

This function is a wrapper around **phytools** getDescendants (*Revell 2012*). It returns the node path from a given node or species to the root of the phylogeny.

# Usage

```
getMommy(tree,N)
```

# **Arguments**

tree a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

N the number of node or tip to perform the function on. The function also works

with tip labels.

#### Value

The function produces a vector of node numbers as integers, collated from a node or a tip towards the tree root.

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

### References

Revell, L. J. (2012). phytools: An R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution*, 3: 217-223.doi:10.1111/j.2041-210X.2011.00169.x

```
data("DataApes")
DataApes$Tstage->Tstage
getMommy(tree=Tstage, N=12)
```

getSis 21

getSis	Get sister clade	

# Description

The function identifies and returns the sister clade of a given node/tip.

# Usage

```
getSis(tree,n,printZoom=TRUE)
```

# Arguments

tree a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

n number of focal node or name of focal tip.

printZoom if TRUE the function plots the tree section of interest.

# Value

The sister node number or sister tip name. In case of polytomies, the function returns a vector.

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

# **Examples**

```
data(DataOrnithodirans)
DataOrnithodirans$treedino->treedino
getSis(tree=treedino,n=677,printZoom=FALSE)
getSis(tree=treedino,n="Shenzhoupterus_chaoyangensis",printZoom=FALSE)
```

lollipoPlot Lollipop charts

# **Description**

The function generates lollipop or dumbbell dots charts.

# Usage

```
lollipoPlot(values, type = "v", pt.lwd = NULL, pt.col = NULL, ...)
```

22 makeFossil

# **Arguments**

values	either a vector, matrix, or data.frame of data. If matrix or data.frame including two columns, a dumbbell dots chart is plotted.
type	plot direction, either vertical ("v", the default) or horizontal ("h").
pt.lwd	points lwd
pt.col	points color
	other arguments passed to the functions plot, points, and segments.

#### **Details**

If a dumbbell dots chart is plotted, different parameters (i.e. col/cex/pch/bg/lwd) for starting and ending points can be supplied. See example for further details.

# Author(s)

Silvia Castiglione, Carmela Serio, Pasquale Raia

# **Examples**

makeFossil

Make fossil species on a phylogeny

# Description

This function takes an object of class 'phylo' and randomly changes the lengths of the leaves.

makeL 23

# Usage

```
makeFossil(tree,p=0.5,ex=0.5)
```

# **Arguments**

tree	a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.
p	the proportion of tips involved. By default it is half of the number of tips.
ex	the multiplying parameter to change the leaf lengths. It is set at 0.5 by default.

#### Value

The function produces a phylogeny having the same backbone of the original one.

#### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

# **Examples**

```
data("DataApes")
DataApes$Tstage->Tstage
makeFossil(tree=Tstage)
```

makeL

Matrix of branch lengths along root-to-tip paths

# Description

This function produces a n \* m matrix, where n=number of tips and m=number of branches (i.e. n + number of nodes). Each row represents the branch lengths aligned along a root-to-tip path.

# Usage

```
makeL(tree)
```

# Arguments

tree

a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

# Value

The function returns a n \* m matrix of branch lengths for all root-to-tip paths in the tree (one per species).

24 makeL1

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

# **Examples**

```
data("DataApes")
DataApes$Tstage->Tstage
makeL(tree=Tstage)
```

makeL1

Matrix of branch lengths along a root-to-node path

# Description

This function produces a n \* n matrix, where n=number of internal branches. Each row represents the branch lengths aligned along a root-to-node path.

# Usage

```
makeL1(tree)
```

# **Arguments**

tree

a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

#### Value

The function returns a n \* n matrix of branch lengths for all root-to-node paths (one per each node of the tree).

### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

```
data("DataApes")
DataApes$Tstage->Tstage
makeL1(tree=Tstage)
```

namesCompare 25

namesCompare

Checking species names for misspelling and synonyms

# Description

The function cross-references two vectors of species names checking for possible synonyms, misspelled names, and genus-species or species-subspecies correspondence.

### Usage

```
namesCompare(vec1, vec2)
```

# **Arguments**

vec1, vec2

a vector of species names. Genus names only are also allowed. Generic name and specific epithet must be separated by '\_'. Note that vec2 is used as the reference. Incomplete or suspicious names are better placed in vec1 (see example below).

#### Value

The function returns a list including:

**\$genus** if vec1 includes genera names which miss specific epithet, this object lists all the species in vec2 belonging to each of the genera.

**\$subspecies** if vec1 includes subspecies (i.e. two epithets after genus name), this object lists species in vec2 possibly corresponding to each of the subspecies.

**\$epithet** lists species with matching epithets as possible synonyms.

**\$misspelling** lists possible misspelled names. For each proposed mismatched names pair the proportion of characters in the vec1 differing from the string in vec2 is returned.

# Author(s)

Silvia Castiglione, Carmela Serio, Antonella Esposito

node.paths

Tracing nodes along paths

# Description

Given a vector of nodes, the function collates nodes along individual lineages from the youngest (i.e. furthest from the tree root) to the oldest.

### Usage

```
node.paths(tree, vec)
```

# **Arguments**

tree a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

vec a vector of node numbers

#### Value

A list of node paths, each starting from the youngest node (i.e. furthest from the tree root) and ending to the oldest along the path.

# Author(s)

Silvia Castiglione, Pasquale Raia

# Examples

```
require(ape)
rtree(100)->tree
sample(seq(Ntip(tree)+1,Ntip(tree)+Nnode(tree)),20)->nods
plot(tree,show.tip.label=FALSE)
nodelabels(node=nods,frame="n",col="red")
node.paths(tree=tree, vec=nods)
```

overfitRR

Testing RRphylo methods overfit

# Description

Testing the robustness of search. trend (Castiglione et al. 2019a), search. shift (Castiglione et al. 2018), search.conv (Castiglione et al. 2019b), and PGLS\_fossil results to sampling effects and phylogenetic uncertainty.

# Usage

```
overfitRR(RR,y,phylo.list=NULL,s=0.25,swap.args=NULL,trend.args=NULL,shift.args=NULL,conv.args=NULL,pgls.args=NULL,aces=NULL,aces.x1=NULL,cov=NULL,rootV=NULL,nsim=100,clus=.5)\\
```

# Arguments

rguments	
RR	an object produced by RRphylo.
У	a named vector of phenotypes.
phylo.list	a list (or multiPhylo) of alternative phylogenies to be tested.
S	the percentage of tips to be cut off. It is set at 25% by default. If phylo.list is provided, this argument is ignored.
swap.args	a list of arguments to be passed to the function <pre>swapONE</pre> , including list(si=NULL, si2=NULL, node=NULL). If swap.arg is unspecified, the function automatically sets both si and si2 to 0.1. If phylo.list is provided, swapping is not performed.
trend.args	a list of arguments specific to the function search.trend, including list(node=NULL,x1.residuals=FA If a trend for the whole tree is to be tested, type trend.args = list(). No trend is tested if left unspecified.
shift.args	a list of arguments specific to the function search.shift, including list(node=NULL, state=NULL). Arguments node and state can be specified at the same time.
conv.args	a list of arguments specific to the function search.conv, including list(node=NULL, state=NULL, declust=FALSE). Arguments node and state can be specified at the same time.
pgls.args	a list of arguments specific to the function PGLS_fossil, including list(modform, data, tree=FALSE, RR=TRUE,). If tree=TRUE, PGLS_fossil is performed by using the RRphylo output tree as tree argument. If RR=TRUE, PGLS_fossil is performed by using the RRphylo output as RR argument. Arguments tree and RR can be TRUE at the same time are further argument passed to PGLS_fossil.
aces	if used to produce the RR object, the vector of those ancestral character values at nodes known in advance must be specified. Names correspond to the nodes in the tree.
x1	the additional predictor to be specified if the RR object has been created using an additional predictor (i.e. multiple version of RRphylo). 'x1' vector must be as long as the number of nodes plus the number of tips of the tree, which can be obtained by running RRphylo on the predictor as well, and taking the vector of ancestral states and tip values to form the x1.
aces.x1	a named vector of ancestral character values at nodes for x1. It must be indicated if the RR object has been created using both aces and x1. Names correspond to the nodes in the tree.
cov	if used to produce the RR object, the covariate must be specified. As in RRphylo, the covariate vector must be as long as the number of nodes plus the number of tips of the tree, which can be obtained by running RRphylo on the covariate as

well, and taking the vector of ancestral states and tip values to form the covariate.

rootV if used to produce the RR object, the phenotypic value at the tree root must be

specified.

nsim number of simulations to be performed. It is set at 100 by default.

clus the proportion of clusters to be used in parallel computing. To run the single-

threaded version of overfitRR set clus = 0.

#### **Details**

Methods using a large number of parameters risk being overfit. This usually translates in poor fitting with data and trees other than the those originally used. With RRphylo methods this risk is usually very low. However, the user can assess how robust the results got by applying search.shift, search.trend, search.conv or PGLS\_fossil are by running overfitRR. With the latter, the original tree and data are subsampled by specifying a s parameter, that is the proportion of tips to be removed from the tree. In some cases, though, removing as many tips as imposed by s would delete too many tips right in clades and/or states under testing. In these cases, the function maintains no less than 5 species at least in each clade/state under testing (or all species if there is less), reducing the sampling parameter s if necessary. Internally, overfitRR further shuffles the tree by using the function swapONE. Thereby, both the potential for overfit and phylogenetic uncertainty are accounted for straight away.

Otherwise, a list of alternative phylogenies can be supplied to overfitRR. In this case subsampling and swapping arguments are ignored, and robustness testing is performed on the alternative topologies as they are. If a clade has to be tested either in search.shift, search.trend, or search.conv, the function scans each alternative topology searching for the corresponding clade. If the species within such clade on the alternative topology differ more than 10 species within the clade in the original tree, the identity of the clade is considered disrupted and the test is not performed.

# Value

The function returns a 'RRphyloList' object containing:

**\$mean.sampling** the mean proportion of species actually removed from the tree over the iterations.

\$tree.list a 'multiPhylo' list including the trees generated within overfitRR

**\$RR.list** a 'RRphyloList' including the results of each RRphylo performed within overfitRR

**\$rootCI** the 95% confidence interval around the root value.

**\$ace.regressions** a 'RRphyloList' including the results of linear regression between ancestral state estimates before and after the subsampling.

**\$conv.results** a list including results for search.conv performed under clade and state conditions. If a node pair is specified within conv.args, the \$clade object contains the percentage of simulations producing significant p-values for convergence between the clades, and the proportion of tested trees (i.e. where the clades identity was preserved; always 1 if no phylo.list is supplied). If a state vector is supplied within conv.args, the object \$state contains the percentage of simulations producing significant p-values for convergence within (single state) or between states (multiple states).

**\$shift.results** a list including results for search.shift performed under clade and sparse conditions. If one or more nodes are specified within shift.args, the \$clade object contains for each node the percentage of simulations producing significant p-value separated by shift sign, and the

same figures by considering all the specified nodes as evolving under a single rate (all.clades). For each node the proportion of tested trees (i.e. where the clade identity was preserved; always 1 if no phylo.list is supplied) is also indicated. If a state vector is supplied within shift.args, the object \$sparse contains the percentage of simulations producing significant p-value separated by shift sign (\$p.states).

**\$trend.results** a list including the percentage of simulations showing significant p-values for phenotypes versus age and absolute rates versus age regressions for the entire tree separated by slope sign (\$tree). If one or more nodes are specified within trend.args, the list also includes the same results at nodes (\$node) and the results for comparison between nodes (\$comparison). For each node the proportion of tested trees (i.e. where the clade identity was preserved; always 1 if no phylo.list is supplied) is also indicated.

**\$pgls.results** two 'RRphyloList' objects including results of PGLS\_fossil performed by using the phylogeny as it is (\$tree) or rescaled according to the RRphylo rates (\$RR).

#### Author(s)

Silvia Castiglione, Carmela Serio, Pasquale Raia

#### References

Castiglione, S., Tesone, G., Piccolo, M., Melchionna, M., Mondanaro, A., Serio, C., Di Febbraro, M., & Raia, P. (2018). A new method for testing evolutionary rate variation and shifts in phenotypic evolution. *Methods in Ecology and Evolution*, 9: 974-983.doi:10.1111/2041-210X.12954

Castiglione, S., Serio, C., Mondanaro, A., Di Febbraro, M., Profico, A., Girardi, G., & Raia, P. (2019a) Simultaneous detection of macroevolutionary patterns in phenotypic means and rate of change with and within phylogenetic trees including extinct species. *PLoS ONE*, 14: e0210101. https://doi.org/10.1371/journal.pone.0210101

Castiglione, S., Serio, C., Tamagnini, D., Melchionna, M., Mondanaro, A., Di Febbraro, M., Profico, A., Piras, P.,Barattolo, F., & Raia, P. (2019b). A new, fast method to search for morphological convergence with shape data. *PLoS ONE*, 14, e0226949. https://doi.org/10.1371/journal.pone.0226949

#### See Also

overfitRR vignette; search.trend vignette; search.shift vignette; search.conv vignette;

```
## Not run:
data("DataOrnithodirans")
DataOrnithodirans$treedino->treedino
DataOrnithodirans$massdino->massdino
DataOrnithodirans$statedino->statedino
cc<- 2/parallel::detectCores()

# Extract Pterosaurs tree and data
library(ape)
extract.clade(treedino,746)->treeptero
massdino[match(treeptero$tip.label,names(massdino))]->massptero
massptero[match(treeptero$tip.label,names(massptero))]->massptero
```

```
RRphylo(tree=treedino,y=massdino)->dinoRates
RRphylo(tree=treeptero,y=log(massptero))->RRptero
# Case 1 search.shift under both "clade" and "sparse" condition
search.shift(RR=dinoRates, status.type= "clade")->SSnode
search.shift(RR=dinoRates, status.type= "sparse", state=statedino)->SSstate
overfitRR(RR=dinoRates,y=massdino,swap.args =list(si=0.2,si2=0.2),
          shift.args = list(node=rownames(SSnode$single.clades), state=statedino),
          nsim=10,clus=cc)->orr.ss
# Case 2 search.trend on the entire tree
search.trend(RR=RRptero, y=log(massptero),nsim=100,clus=cc,cov=NULL,node=NULL)->STtree
overfitRR(RR=RRptero,y=log(massptero),swap.args =list(si=0.2,si2=0.2),
          trend.args = list(),nsim=10,clus=cc)->orr.st1
# Case 3 search.trend at specified nodescov=NULL,
search.trend(RR=RRptero, y=log(massptero),node=143,clus=cc)->STnode
overfitRR(RR=RRptero,y=log(massptero),
          trend.args = list(node=143),nsim=10,clus=cc)->orr.st2
# Case 4 overfitRR on multiple RRphylo
data("DataCetaceans")
DataCetaceans$treecet->treecet
DataCetaceans$masscet->masscet
DataCetaceans$brainmasscet->brainmasscet
DataCetaceans$aceMyst->aceMyst
ape::drop.tip(treecet, treecet$tip.label[-match(names(brainmasscet),
                                               treecet$tip.label)])->treecet.multi
masscet[match(treecet.multi$tip.label,names(masscet))]->masscet.multi
RRphylo(tree=treecet.multi,y=masscet.multi)->RRmass.multi
RRmass.multi$aces[,1]->acemass.multi
c(acemass.multi,masscet.multi)->x1.mass
RRphylo(tree=treecet.multi,y=brainmasscet,x1=x1.mass)->RRmulti
search.trend(RR=RRmulti, y=brainmasscet,x1=x1.mass,clus=cc)->STcet
overfitRR(RR=RRmulti,y=brainmasscet,trend.args = list(),
          x1=x1.mass,nsim=10,clus=cc)->orr.st3
search.trend(RR=RRmulti, y=brainmasscet,x1=x1.mass,x1.residuals=TRUE,
             clus=cc)->STcet.resi
overfitRR(RR=RRmulti,y=brainmasscet,trend.args = list(x1.residuals=TRUE),
          x1=x1.mass,nsim=10,clus=cc)->orr.st4
# Case 5 searching convergence between clades and within a single state
data("DataFelids")
DataFelids$PCscoresfel->PCscoresfel
```

PGLS\_fossil 31

```
DataFelids$treefel->treefel
 DataFelids$statefel->statefel
 RRphylo(tree=treefel,y=PCscoresfel,clus=cc)->RRfel
 search.conv(RR=RRfel, y=PCscoresfel, min.dim=5, min.dist="node9",clus=cc)->SC.clade
 as.numeric(c(rownames(SC.clade[[1]])[1], as.numeric(as.character(SC.clade[[1]][1,1])))) -> conv.nodes
 overfitRR(RR=RRfel, y=PCscoresfel,conv.args =
 list(node=conv.nodes,state=statefel,declust=TRUE),nsim=10,clus=cc)->orr.sc
 # Case 6 overfitRR on PGLS_fossil
 library(phytools)
 rtree(100)->tree
 fastBM(tree)->resp
 fastBM(tree,nsim=3)->resp.multi
 fastBM(tree)->pred1
 fastBM(tree)->pred2
 PGLS_fossil(modform=y1~x1+x2,data=list(y1=resp,x2=pred1,x1=pred2),tree=tree)->pgls_noRR
 RRphylo::RRphylo(tree,resp)->RR
 PGLS\_fossil(modform=y1~x1+x2,data=list(y1=resp,x2=pred1,x1=pred2),tree=tree,RR=RR)->pgls\_RR
 overfitRR(RR=RR,y=resp,
           pgls.args=list(modform=y1~x1+x2,data=list(y1=resp,x2=pred1,x1=pred2),
                           tree=TRUE,RR=TRUE),nsim=10,clus=cc)->orr.pgls1
 PGLS_fossil(modform=y1~x1+x2,data=list(y1=resp.multi,x2=pred1,x1=pred2),tree=tree)->pgls2_noRR
 cc<- 2/parallel::detectCores()</pre>
 RRphylo::RRphylo(tree,resp.multi,clus=cc)->RR
 PGLS_fossil(modform=y1~x1+x2,data=list(y1=resp.multi,x2=pred1,x1=pred2),tree=tree,RR=RR)->pgls2_RR
 overfitRR(RR=RR,y=resp.multi,
           pgls.args=list(modform=y1~x1+x2,data=list(y1=resp.multi,x2=pred1,x1=pred2),
                           tree=TRUE,RR=TRUE),nsim=10,clus=cc)->orr.pgls2
 ## End(Not run)
PGLS_fossil
                         Phylogenetic Generalized Least Square with phylogenies including
                         fossils
```

# **Description**

The function performs pgls for non-ultrametric trees using a variety of evolutionary models or RRphylo rates to change the tree correlation structure.

32 PGLS\_fossil

# Usage

```
PGLS_fossil(modform,data,tree=NULL,RR=NULL,...)
```

# **Arguments**

modform the formula for the regression model.

data a list of named vectors including response and predictor variables as named in modform.

tree a phylogenetic tree to be indicated for any model other than RRphylo is used to rescale tree branches. The tree needs not to be ultrametric and fully dichotomous.

RR the result of RRphylo performed on the response variable. If RR is specified, tree branches are rescaled to the absolute branch-wise rate values calculated by RRphylo to transform the variance-covariance matrix.

... further argument passed to the function phylolm.

#### Value

Fitted pgls parameters and significance.

# Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

# See Also

RRphylo vignette

```
## Not run:
library(ape)
library(phytools)

rtree(100)->tree
fastBM(tree)->resp
fastBM(tree,nsim=3)->resp.multi
fastBM(tree)->pred1
fastBM(tree)->pred2

PGLS_fossil(modform=y1~x1+x2,data=list(y1=resp,x2=pred1,x1=pred2),tree=tree)->pgls_noRR

RRphylo::RRphylo(tree,resp)->RR
PGLS_fossil(modform=y1~x1+x2,data=list(y1=resp,x2=pred1,x1=pred2),tree=tree,RR=RR)->pgls_RR

# To derive log-likelihood and AIC for PGLS_fossil outputs the function AIC can be applied
AIC(pgls_noRR)
AIC(pgls_RR)
```

phyloclust 33

```
PGLS_fossil(modform=y1~x1+x2,data=list(y1=resp.multi,x2=pred1,x1=pred2),tree=tree)->pgls2_noRR
cc<- 2/parallel::detectCores()
RRphylo::RRphylo(tree,resp.multi,clus=cc)->RR
PGLS_fossil(modform=y1~x1+x2,data=list(y1=resp.multi,x2=pred1,x1=pred2),tree=tree,RR=RR)->pgls2_RR
AIC(pgls2_noRR)
AIC(pgls2_RR)
## End(Not run)
```

phyloclust

Test for phylogenetic clustering

# **Description**

The function tests the presence of phylogenetic clustering for species within a focal state.

### Usage

```
phyloclust(tree, state, focal, nsim=100)
```

# **Arguments**

tree a phylogenetic tree. The tree needs not to be ultrametric or fully dichotomous.

state the named vector of tip states.

focal the focal state to be tested for phylogenetic clustering.

nsim number of simulations to perform the phylogenetic clustering test.

# **Details**

To test for phylogenetic clustering, the function computes the mean cophenetic (i.e. evolutionary time) distance between all the species under the focal state. Such value is compared to a random distribution of time distances obtained by sampling nsim times as many random tips as those under the focal state. In the presence of significant phylogenetic clustering, tips under the focal state are randomly removed until the p becomes >0.05 or only 3 tips are left.

### Value

The function returns a list including the p-value (\$p) for the test of phylogenetic clustering and a \$declusterized object containing the declusterized versions of the original tree and state vector (i.e. tips are removed as to make p>0.05) and the vector of removed species.

# Author(s)

Silvia Castiglione, Pasquale Raia

34 plotConv

# **Examples**

```
data("DataFelids")
DataFelids$treefel->treefel
DataFelids$statefel->statefel
phyloclust(tree=treefel,state=statefel,focal="saber")
```

plotConv

Graphical representation of search.conv results

# **Description**

This function generates customized functions to produce plots out of search.conv results.

# Usage

```
plotConv(SC,y,variable,RR=NULL,state=NULL,aceV=NULL)
```

# **Arguments**

SC an object produced by search.conv.

y the multivariate phenotype used to perform search.conv.

variable the index of result to plot. If convergence between clades is inspected, this is

the position within the SC\$'average distance from group centroids' vector of the clade pair to be plotted. In the case of convergence between states, this is the number of the line of SC\$state.res where results for the state pair are

returned.

RR the object produced by RRphylo used to perform search.conv. This is not

indicated if convergence between states is tested.

state the named vector of tip states used to perform search.conv. This is not indi-

cated if convergence between clades is tested.

aceV phenotypic values at internal nodes to be provided if used to perform search.conv.

# Value

If convergence between clades was tested, plotConv returns a list of four functions:

\$plotHistTips shows the mean Euclidean distance computed between phenotypic vectors of all
the tips belonging to the converging clades as compared to the distribution of distances between all
possible pair of tips across the rest of the tree. The usage is: ...\$plotHistTips(hist.args=NULL,line.args=NULL),
where hist.args is a list of further arguments passed to the function hist, and line.args is a list
of further arguments passed to the function lines.

\$plotHistAces shows the Euclidean distance computed between phenotypic vectors of the MR-CAs of the converging clades as compared to the distribution of distances between all possible pairs of nodes across the rest of the tree. The usage is identical to \$plotHistTips.

plotConv 35

\$plotPChull generates a PC1/PC2 plot obtained by performing a PCA of the species phenotypes. Convergent clades are indicated by colored convex hulls. Large dots represent the mean phenotypes per clade (i.e. their group centroids) and asterisks (customizable) represent the ancestral phenotypes of the individual clades. The usage is: ...\$plotPChull(plot.args=NULL,chull.args=NULL,means.args=NULL, ace.args=NULL,legend.args=list(),where plot.args is a list of further arguments passed to the function plot, chull.args is a list of further arguments passed to the function polygon, means.args and ace.args are lists of further argument passed to the function points to customize the dots representing the centroids and the ancestral phenotypes respectively, and legend.args is a list of additional arguments passed to the function legend (if = NULL the legend is not plotted).

\$plotTraitgram produces a modified traitgram plot (see package **picante**) highlighting the branches of the clades found to converge. The usage is: plotTraitgram(colTree=NULL,colNodes=NULL,...), where colTree is the color to represent the traitgram lines not pertaining the converging clades, colNodes is the color (or the vector of colors) to represent the traitgram lines pertaining the converging clades, and ... are further arguments passed to the function plot to plot lines.

If convergence between states was tested, plotConv returns a list of two functions:

\$plotPChull generates a PC1/PC2 plot obtained by performing a PCA of the species phenotypes, with colored convex hulls enclosing species belonging to different states. The usage is: ...\$plotPChull(plot.args=NULL,chull.args=NULL,points.args=NULL,legend.args=list(), where plot.args is a list of further arguments passed to the function plot, chull.args is a list of further arguments passed to the function polygon, points.args is a list of further argument passed to the function points, and legend.args is a list of additional arguments passed to the function legend (if = NULL the legend is not plotted).

\$plotPolar produces a polar plot of the mean angle within/between state/s as compared to the 95 angles. The usage is: ...\$plotPolar(polar.args=NULL,polygon.args=NULL,line.args=NULL), where polar.args is a list of further arguments passed to the function polar.plot to set the plot basics (i.e. radial.lim, start, and so on), polygon.args is a list of further arguments passed to the function polar.plot under the condition rp.type="p" (see plotrix::polar.plot for details) to set the angles distribution graphics, and line.args is a list of further arguments passed to the function polar.plot under the condition rp.type="r" to set the mean angle graphics.

#### Author(s)

Silvia Castiglione

#### References

Castiglione, S., Serio, C., Tamagnini, D., Melchionna, M., Mondanaro, A., Di Febbraro, M., Profico, A., Piras, P.,Barattolo, F., & Raia, P. (2019). A new, fast method to search for morphological convergence with shape data. *PLoS ONE*, 14, e0226949. https://doi.org/10.1371/journal.pone.0226949

# See Also

search.conv vignette
plotConv vignette

36 plotRates

# **Examples**

```
## Not run:
data("DataFelids")
DataFelids$PCscoresfel->PCscoresfel
DataFelids$treefel->treefel
DataFelids$statefel->statefel->state2
state2[sample(which(statefel=="nostate"),20)]<-"st2"
cc<- 2/parallel::detectCores()</pre>
RRphylo(treefel, PCscoresfel, clus=cc)->RRfel
search.conv(RR=RRfel, y=PCscoresfel, min.dim=5, min.dist="node9",clus=cc)->sc_clade
plotConv(sc_clade,PCscoresfel,variable=2,RR=RRfel)->pc
pc$plotHistTips(hist.args = list(col="gray80",yaxt="n",cex.axis=0.8,cex.main=1.5),
                line.args = list(lwd=3,lty=4,col="purple"))
pc$plotHistAces(hist.args = list(col="gray80",cex.axis=0.8,cex.main=1.5),
                line.args = list(lwd=3,lty=4,col="gold"))
pc$plotPChull(chull.args = list(border=c("cyan", "magenta"), lty=1),
              means.args = list(pch=c(23,22),cex=3,bg=c("cyan2","magenta2")),
              ace.args=list(pch=9),legend.args = NULL)
pc$plotTraitgram(colTree = "gray70",colNodes = c("cyan","magenta"))
search.conv(tree=treefel, y=PCscoresfel, state=statefel,declust=TRUE,clus=cc)->sc_state
plotConv(sc_state,PCscoresfel,variable=1,state=statefel)->pc
pc$plotPChull(chull.args = list(border=c("gray70","blue"),lty=1),
              points.args = list(pch=c(23,22),bg="gray"),
              legend.args = list(pch=c(23,22),x="top"))
pc$plotPolar(polar.args = list(clockwise=TRUE,start=0,rad.col="black",grid.col="black"),
             polygon.args = list(line.col="green",poly.col=NA,lwd=2),
             line.args = list(line.col="deeppink",lty=2,lwd=3))
## End(Not run)
```

plotRates

Plot RRphylo rates at a specified node

# **Description**

This function generates customized functions to produce histograms and lollipop charts of the RRphylo rates computed for a given clade as compared to the rates computed for the rest of the tree.

# Usage

```
plotRates(RR, node, export.tiff=NULL, filename=NULL)
```

plotRates 37

#### **Arguments**

RR an object produced by RRphylo.

node the node subtending the clade of interest.

export.tiff is deprecated. filename is deprecated.

#### Value

The function returns a list of functions:

**\$plotHist** returns the histograms of rates (in ln absolute values) computed for the focal clade against rates computed the rest of the tree. The usage is: ...\$plotHist(hist.args=list(col1,col2),legend.args=list()), where legend.args is a list of additional arguments passed to the function legend (if = NULL the legend is not plotted) and hist.args is a list of further arguments passed to the function plot.histogram. hist.args default arguments include histogram colors for background rates (col1) and rates of the clade under inspection (col2).

**\$plotLollipop** returns the lollipop chart of the rates of individual branches of the focal clade collated in increasing rate value, and contrasted to the average rate computed over the rest of the tree branches (the vertical line). The usage is: ...\$plotLollipop(lollipop.args=NULL,line.args=NULL), where lollipop.args is a list of further arguments passed to the function lollipoPlot and line.args is a list of additional arguments passed to the function line. This function additionally returns the vector of rates for the focal clade, collated in increasing order.

#### Author(s)

Silvia Castiglione, Pasquale Raia

#### See Also

```
RRphylo vignette
RRphylo vignette
plotRates vignette
```

38 plotShift

plotShift	Graphical representation of search.shift results
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# **Description**

This function generates customized functions to produce plots out of search.shift results.

## Usage

```
plotShift(RR,SS,state=NULL)
```

#### **Arguments**

RR the object produced by RRphylo used to perform search.shift.

SS an object produced by search.shift.

state if search.shift was performed under status.type = "sparse", this is the

same state vector provided to the function.

#### Value

The function returns a function to generate a customizable plot of search.shift results.

If search.shift was performed under status.type = "clade", plotShift returns a \$plotClades function which highlights the shifting clades onto the phylogenetic tree. The usage is: ...\$plotClades(tree.args=NULL, s) where tree.args is a list of further arguments passed to the function plot.phylo, and symbols.args is a list of further arguments passed to the function symbols (n.b. the shape of the symbol is not customizable). The function automatically plots red circles for negative shifts and blue circles for positive shifts, in each cases with the radium proportional to the absolute value of rate difference. The user can choose different color options for positive/negative shifts by setting symbols.args=list(fg=c(pos="color for positive shift", neg="color for negative shift")), or provide a vector of as many colors as the number of shifting clades. The same applies to the argument "bg".

If search.shift was performed under status.type = "sparse", plotShift returns a \$plotStates function which plots the states onto the phylogenetic trees. The usage is: ...\$plotStates(tree.args=NULL,points.args=where tree.args is a list of further arguments passed to the function plot.phylo, points.args is a list of further arguments passed to the function points, and legend.args is a list of additional arguments passed to the function legend (if = NULL the legend is not plotted). If as many colors/pch values as the number of different states are provided in points.args, each of them is assigned to each states taken in the same order they are returned by search.shift.

#### Author(s)

Silvia Castiglione

#### References

Castiglione, S., Tesone, G., Piccolo, M., Melchionna, M., Mondanaro, A., Serio, C., Di Febbraro, M., & Raia, P.(2018). A new method for testing evolutionary rate variation and shifts in phenotypic evolution. *Methods in Ecology and Evolution*, 9: 974-983.doi:10.1111/2041-210X.12954

plotTrend 39

## See Also

```
search.shift vignette plotShift vignette
```

# **Examples**

```
## Not run:
data("DataOrnithodirans")
DataOrnithodirans$treedino->treedino
DataOrnithodirans$massdino->massdino
DataOrnithodirans$statedino->statedino
RRphylo(tree=treedino,y=massdino)->dinoRates
search.shift(RR=dinoRates, status.type="clade")->SSauto
plotShift(RR=dinoRates,SS=SSauto)->plotSS
plotSS$plotClades()
search.shift(RR=dinoRates, status.type="clade", node=c(696,746))->SSnode
plotShift(RR=dinoRates,SS=SSnode)->plotSS
plotSS$plotClades(tree.args=list(no.margin=TRUE),
                  symbols.args=list(fg=NA,bg=c(pos="cyan",neg="magenta")))
search.shift(RR=dinoRates, status.type= "sparse", state=statedino)->SSstate
plotShift(RR=dinoRates,SS=SSstate,state=statedino)->plotSS
plotSS$plotStates(tree.args=list(no.margin=TRUE),
                  points.args=list(bg=c("gold","forestgreen","royalblue","white"),
                                   col=c("black","black","black","orangered"),
                                   pch=c(21,22,24,11)),legend.args=list())
## End(Not run)
```

plotTrend

Graphical representation of search.trend results

# Description

This function generates customized functions to produce plots of phenotype versus time and absolute evolutionary rates versus time regressions for the entire phylogeny and individual clades. Each custom function takes as first argument the index or name of the variable (as in the search.trend output in \$trends.data\$phenotypeVStime) to be plotted.

# Usage

```
plotTrend(ST)
```

## **Arguments**

ST

an object produced by search. trend.

40 plotTrend

#### Value

The function returns a list of functions:

\$plotSTphen returns the plot of rescaled phenotype versus age regression. The 95% confidence intervals of slopes produced by regressing phenotypes simulated under the Brownian motion are plotted as a polygon. The usage is . . . \$plotSTphen(variable,plot.args=NULL,polygon.args=NULL,line.args=NULL), where variable is the index or name of the variable to be plotted, plot.args is a list of further arguments passed to the function plot, polygon.args is a list of further arguments passed to the function polygon, and line.args is a list of further arguments passed to the function lines. The functions automatically plots white points for internal nodes, red points for tips, a blue regression line, and a gray shaded polygon to represent the 95% confidence intervals of Brownian motion slopes.

\$plotSTrates returns the plot of log rescaled rates versus age regression. The 95% confidence intervals of slopes produced by regressing rates simulated under the Brownian motion are plotted as a shaded area. The arguments are the same as described for \$plotSTphen. In the case of multivariate y, the 2-norm vector of multiple rates (see RRphylo for details) can be plotted by setting variable = "rate" or variable = the number of actual variables + 1.

\$plotSTphenNode returns plots of rescaled phenotype versus age regression for individual clades. The usage is . . . \$plotSTphenNode(variable, node, plot.args=NULL, lineTree.args=NULL, lineNode.args=NULL, node where variable is the same as plotSTphen and node is the vector of indices or numbers (as inputted to search.trend, to be indicated as character) of nodes to be plotted. The function allows up to nine nodes at the same time. plot.args is a list of further arguments passed to the function plot, including pch.node, a custom argument to set individual pch at nodes (its length can be one or as long as the number of nodes). lineTree.args is a list of further arguments passed to the function lines used to plot the regression line for the entire tree lineNode.args is a list of further arguments passed to the function lines used to plot the regression line for individual nodes. node.palette is the vector of colors specific to nodes points and lines. Its length can be one or as long as the number of nodes.

\$plotSTratesNode returns plots of absolute rates versus age regression for individual clades. The arguments are the same as described for \$plotSTphenNode. In the case of multivariate y, the 2-norm vector of multiple rates (see RRphylo for details) can be plotted by setting variable = "rate" or variable = the number of actual variables + 1.

#### Author(s)

Silvia Castiglione, Carmela Serio

#### References

Castiglione, S., Serio, C., Mondanaro, A., Di Febbraro, M., Profico, A., Girardi, G., & Raia, P. (2019) Simultaneous detection of macroevolutionary patterns in phenotypic means and rate of change with and within phylogenetic trees including extinct species. *PLoS ONE*, 14: e0210101. https://doi.org/10.1371/journal.pone.0210101

#### See Also

search.trend vignette
plotTrend vignette

random.evolvability.test

#### **Examples**

```
## Not run:
data("DataOrnithodirans")
DataOrnithodirans$treedino->treedino
DataOrnithodirans$massdino->massdino
cc<- 2/parallel::detectCores()</pre>
# Extract Pterosaurs tree and data
library(ape)
extract.clade(treedino,746)->treeptero
massdino[match(treeptero$tip.label,names(massdino))]->massptero
massptero[match(treeptero$tip.label,names(massptero))]->massptero
RRphylo(tree=treeptero,y=log(massptero))->RRptero
search.trend(RR=RRptero, y=log(massptero), nsim=100, node=143, clus=cc,cov=NULL)->ST
plotTrend(ST)->plotST
plotST$plotSTphen(1) # to plot phenotypic trend through time for entire tree
plotST$plotSTphen("log(massptero)",plot.args=list(cex=1.2,col="blue")) # equivalent to above
plotST$plotSTrates(1) # to plot rates trend through time for entire tree
# to plot phenotypic trend through time for the clade
plotST$plotSTphenNode("log(massptero)",plot.args=list(xlab="Age",main="Phenotypic trend"),
                    lineTree.args=list(lty=1,col="pink"),lineNode.args=list(lwd=3),
                    node.palette=c("chartreuse"))
# to plot rates trend through time for the clade
plotST$plotSTratesNode("rate")
## End(Not run)
```

random.evolvability.test

Randomization test for phylogenetic structuring in evolvability

# **Description**

The function is a wrapper around the function MeanMatrixStatistics from the package evolqg (Melo et al. 2015). It estimates ancestral character at internal nodes either according to Brownian Motion or by means of RRphylo (see the argument node.estimation), then performs MeanMatrixStatistics to calculate: Mean Squared Correlation, ICV, Autonomy, ConditionalEvolvability, Constraints, Evolvability, Flexibility, Pc1Percent, and Respondability. To assess the importance of phylogenetic structuring (signal) on Respondability Evolvability, and Flexibility. The function performs a randomization test by randomly shuffling the species on tree and replicating the analyses nsim times. A p-value is computed by contrasting the real metrics to the ones derived by randomization.

## Usage

```
random.evolvability.test(tree,data,node.estimation=c("RR","BM"),
   aces=NULL,iterations=1000,nsim=100,clus=0.5)
```

#### **Arguments**

tree a phylogenetic tree. The tree needs not to be ultrametric or fully dichotomous.

data a matrix or data.frame of phenotypic data having species as rownames

node.estimation

specify the method to compute ancestral character at nodes. It can be one of "RR", to compute ancestral states by mean of RRphylo, or "BM", to use **phytools**' function fastAnc (*Paradis & Schliep 2019*) to estimate ancestral characters at

nodes according to Brownian Motion.

aces a named matrix of known ancestral character values at nodes. Names correspond

to the nodes in the tree.

iterations the iterations argument to be indicated in MeanMatrixStatistics

nsim the number of simulations to be performed for the randomization test, by default

nrep is set at 100.

clus the proportion of clusters to be used in parallel computing. To run the single-

threaded version of NOME set clus = 0.

#### Value

The function returns a list object including (\$means) the mean values for all statistics as produced by MeanMatrixStatistics and (\$means) the significance levels for Respondability, Evolvability, and Flexibility.

#### Author(s)

Silvia Castiglione, Gabriele Sansalone, Pasquale Raia

#### References

Melo, D., Garcia, G., Hubbe, A., Assis, A. P., & Marroig, G. (2015). EvolQG-An R package for evolutionary quantitative genetics. *F1000Research*, 4.

Revell, L. J. (2012) phytools: An R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution*, 3, 217-223.

## See Also

RRphylo vignette

rate.map 43

#### **Examples**

```
## Not run:
library(ape)
library(phytools)

rtree(30)->tree
fastBM(tree,nsim=4)->y

random.evolvability.test(tree=tree,data=y,node.estimation="RR")->rEvTest
## End(Not run)
```

rate.map

Mapping rate and direction of phenotypic change on 3D surfaces.

# Description

Given vectors of RW (or PC) scores, the function selects the RW(PC) axes linked to highest (and lowest) evolutionary rate values and reconstructs the morphology weighted on them. In this way, rate.map shows where and how the phenotype changed the most between any pair of taxa.

#### Usage

```
rate.map(x, RR, PCscores, pcs, mshape, out.rem = TRUE,
   shape.diff=FALSE, show.names=TRUE)
```

# **Arguments**

x the species/nodes to be compared; it can be a single species, or a vector contain-

ing two species or a species and any of its parental nodes.

RR an object generated by using the RRphylo function

PCscores PC scores.

pcs RW (or PC) vectors (eigenvectors of the covariance matrix) of all the samples.

mshape the Consensus configuration.

out.rem logical: if TRUE mesh triangles with outlying area difference are removed. shape.diff logical: if TRUE, the mesh area differences are displayed in an additional 3d plot.

show. names logical: if TRUE, the names of the species are displayed in the 3d plot.

#### **Details**

After selecting the PC axes, rate.map automatically builds a 3D mesh on the mean shape calculated from the Relative Warp Analysis (RWA) or Principal Component Analysis (PCA) (Schlager 2017) by applying the function vcgBallPivoting (Rvcg). Then, it compares the area differences between corresponding triangles of the 3D surfaces reconstructed for the species and surface of the mrca. Finally, rate.map returns a 3D plot showing such comparisons displayed on shape of the mrca used

44 rate.map

as the reference. The colour gradient goes from blue to red, where blue areas represent expansion of the mesh, while the red areas represent contractions of the mesh triangles. In the calculation of the differences of areas we supply the possibility to find and remove outliers from the vectors of areas calculated on the reference and target surfaces. We suggest considering this possibility if the mesh may contain degenerate facets. Additionally, rate.map allows to investigate the pure morphological comparison of shapes by excluding the evolutionary rate component by setting the argument show.diff = TRUE. In this case, a second 3D plot will be displayed highlighting area differences in terms of expansion (green) and contraction (yellow).

#### Value

The function returns a list including:

- **\$selected** a list of PCs axes selected for higher evolutionary rates for each species.
- **\$surfaces** a list of reconstructed coloured surfaces of the given species and of the most recent common ancestor.

## Author(s)

Marina Melchionna, Antonio Profico, Silvia Castiglione, Gabriele Sansalone, Pasquale Raia

#### References

Schlager, S. (2017). Morpho and Rvcg-Shape Analysis in R: R-Packages for geometric morphometrics, shape analysis and surface manipulations. In: Statistical shape and deformation analysis. Academic Press. Castiglione, S., Melchionna, M., Profico, A., Sansalone, G., Modafferi, M., Mondanaro, A., Wroe, S., Piras, P., & Raia, P. (2021). Human face-off: a new method for mapping evolutionary rates on three-dimensional digital models. Palaeontology. doi:10.1111/pala.12582

#### See Also

```
RRphylo vignette; relWarps; procSym
```

rescaleRR 45

rescaleRR	Rescaling phylogenetic trees	

# Description

The function rescales all branches and leaves of the phylogenetic tree.

# Usage

rescaleRR(tree,RR=NULL,height=NULL,trend=NULL,delta=NULL,kappa=NULL,lambda=NULL)

# Arguments

tree	the phylogenetic tree to be rescaled.
RR	is the output of RRphylo performed on tree. If this parameter is indicated, the tree branches are rescaled according to branch-wise phenotypic evolutionary rates fitted by RRphylo. When a multivariate phenotype is used, rescaling is operated on the norm-2 vector of rates.
height	is the desired height of the tree. If this parameter is indicated, the tree branches are rescaled to match the total height.
trend	is a diffusion model with linear trend in rates through time. The trend scaling is largely based on package <b>geiger</b> 's rescale.phylo function.
delta	if this parameter is indicated, the tree is rescaled according to Pagel's delta transform (Pagel 1999). Nodes are pushed toward the present for values of delta ranging between 0 and 1. The converse applies for delta larger than 1. Negative delta values are not allowed.
kappa	if this parameter is indicated, the tree is rescaled according to Pagel's kappa transform (Pagel 1999). At kappa = 1 the tree is left unmodified. Branches become increasingly closer to 1 as kappa approaches 0, making evolution independent from branch lengths. Negative kappa values are not allowed.
lambda	if this parameter is indicated, the tree is rescaled according to Pagel's lambda transform (Pagel 1999). At lambda = 1 the tree is left unmodified. The tree approaches a star phylogeny as lambda approaches zero. lambda values larger than one are undefined Negative lambda values are not allowed.

# Value

Rescaled phylogenetic tree.

# Author(s)

Silvia Castiglione, Pasquale Raia

#### References

Castiglione, S., Serio, C., Piccolo, M., Mondanaro, A., Melchionna, M., Di Febbraro, M., Sansalone, G., Wroe, S., & Raia, P. (2020). The influence of domestication, insularity and sociality on the tempo and mode of brain size evolution in mammals. *Biological Journal of the Linnean Society*,132: 221-231. doi:10.1093/biolinnean/blaa186

Pagel, M. (1999). Inferring the historical patterns of biological evolution. *Nature*, 401:877-884.

#### **Examples**

```
## Not run:
ape::rtree(100)->tree
phytools::fastBM(tree)->y
max(diag(vcv(tree)))->H

RRphylo::RRphylo(tree,y)->RR
rescaleRR(tree,RR=RR)->treeRR

rescaleRR(tree,height=H/3)->tree_height

rescaleRR(tree,trend=5)->tree_trend

rescaleRR(tree,delta=0.5)->tree_delta05
rescaleRR(tree,delta=2)->tree_delta2

rescaleRR(tree,kappa=0.5)->tree_kappa

rescaleRR(tree,lambda=0.5)->tree_lambda

## End(Not run)
```

retrieve.angles

Extracting a user-specified subset of the evo.dir results

# Description

This function takes the result list produced by evo.dir as the input, and extracts a specific subset of it. The user may specify whether to extract the set of angles between species resultant vectors and the MRCA, the size of resultant vectors, or the set of angles between species.

# Usage

```
retrieve.angles(angles.res,wishlist=c("anglesMRCA","angleDir","angles.between.species"),
  random=c("yes","no"),focus=c("node","species","both","none"),
  node=NULL,species=NULL,write2csv=c("no","yes"),csvfile=NULL)
```

#### **Arguments**

angles.res the object resulting from evo.dir function.

wishlist specifies whether to extract angles and sizes ("anglesMRCA") of resultant vec-

tors between individual species and the MRCA, angles and sizes ("angleDir") of vectors between individual species and a fixed reference vector (the same for

all species), or angles between species resultant vectors ("angles.between.species").

random it needs to be "yes" if 'angles.res' object contains randomization results.

focus it can be "node", "species", "both", or "none", whether the user wants the

results for a focal node, or for a given species, for both, or just wants to visualize

everything.

node must be indicated if focus = "node" or "both". As for evo.dir, the node num-

ber must refer to the dichotomic version of the original tree, as produced by

RRphylo.

species must be indicated if focus = "species" or "both".

write2csv has been deprecated; please see the argument csvfile instead.

csvfile if results should be saved to a .csv file, a character indicating the name of the csv

file and the path where it is to be saved. If no path is indicated the file is stored

in the working directory. If left unspecified, no file will be saved.

#### **Details**

retrieve. angles allows to focalize the extraction to a particular node, species, or both. Otherwise it returns the whole dataset.

# Value

retrieve.angles outputs an object of class 'data.frame'.

If wishlist = "anglesMRCA", the data frame includes:

- MRCA the most recent common ancestor the angle is computed to
- species ID
- angle the angle between the resultant vector of species and the MRCA
- vector.size the size of the resultant vector computed from species to MRCA

If wishlist = "angleDir", the data frame includes:

- MRCA the most recent common ancestor the vector is computed to
- species ID
- angle.direction the angle between the vector of species and a fixed reference
- vector.size the size of the vector of species

If wishlist = "angles.between.species", the data frame includes:

- MRCA the most recent common ancestor the vector is computed from
- species pair IDs of the species pair the "angle between species" is computed for
- angleBTWspecies2MRCA angle between species resultant vectors to MRCA
- anglesBTWspecies angle between species resultant vectors

#### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

```
## Not run:
   data("DataApes")
   DataApes$PCstage->PCstage
   DataApes$Tstage->Tstage
   cc<- 2/parallel::detectCores()</pre>
   RRphylo(tree=Tstage,y=PCstage,clus=cc)->RR
# Case 1. "evo.dir" without performing randomization
   evo.dir(RR,angle.dimension="rates",pair.type="node",
   node= 57,random="no")->evo.p
# Case 1.1 angles and sizes of resultant vectors between individual species and the MRCA:
 # for a focal node
   retrieve.angles(evo.p,wishlist="anglesMRCA",random="no",focus="node",
   node=68)
 # for a focal species
   retrieve.angles(evo.p,wishlist="anglesMRCA",random="no",focus="species",
   species="Sap")
 # for both focal node and species
   retrieve.angles(evo.p,wishlist="anglesMRCA",random="no",focus="both",
   node=68, species="Sap")
 # without any specific requirement
   retrieve.angles(evo.p,wishlist="anglesMRCA",random="no",focus="none")
# Case 1.2 angles and sizes of vectors between individual species
#and a fixed reference vector:
 # for a focal node
   retrieve.angles(evo.p,wishlist="angleDir",random="no",focus="node",
   node=68)
 # for a focal species
   retrieve.angles(evo.p,wishlist="angleDir",random="no",focus="species",
   species="Sap")
 # for both focal node and species
   retrieve.angles(evo.p,wishlist="angleDir",random="no",focus="both",
   node=68, species="Sap")
 # without any specific requirement
   retrieve.angles(evo.p,wishlist="angleDir",random="no",focus="none")
# Case 1.3 angles between species resultant vectors:
 # for a focal node
   retrieve.angles(evo.p,wishlist="angles.between.species",random="no",
   focus="node", node=68)
 # for a focal species
   retrieve.angles(evo.p,wishlist="angles.between.species",random="no",
   focus="species", species="Sap")
```

```
# for both focal node and species
   retrieve.angles(evo.p,wishlist="angles.between.species",random="no",
   focus="both",node=68,species="Sap")
 # without any specific requirement
   retrieve.angles(evo.p,wishlist="angles.between.species",random="no",
   focus="none")
# Case 2. "evo.dir" with performing randomization
   evo.dir(RR, angle.dimension="rates", pair.type="node", node=57,
   random="yes",nrep=10)->evo.p
# Case 2.1 angles and sizes of resultant vectors between individual species and the MRCA:
   retrieve.angles(evo.p,wishlist="anglesMRCA",random="yes",focus="node",
   node=68)
 # for a focal species
   retrieve.angles(evo.p,wishlist="anglesMRCA",random="yes", focus="species",
   species="Sap")
 # for both focal node and species
   retrieve.angles(evo.p,wishlist="anglesMRCA",random="yes",focus="both",
   node=68, species="Sap")
 # without any specific requirement
   retrieve.angles(evo.p,wishlist="anglesMRCA",random="yes",focus="none")
# Case 2.2 angles and sizes of vectors between individual species and a fixed reference vector:
 # for a focal node
   retrieve.angles(evo.p,wishlist="angleDir",random="yes",focus="node",
   node=68)
 # for a focal species
   retrieve.angles(evo.p,wishlist="angleDir",random="yes",focus="species",
   species="Sap")
 # for both focal node and species
   retrieve.angles(evo.p,wishlist="angleDir",random="yes",focus="both",
   node=68, species="Sap")
 # without any specific requirement
   retrieve.angles(evo.p,wishlist="angleDir",random="yes",focus="none")
# Case 2.3 retrieve angles between species resultant vectors:
 # for a focal node
   retrieve.angles(evo.p,wishlist="angles.between.species",random="yes",
   focus="node", node=68)
 # for a focal species
   retrieve.angles(evo.p,wishlist="angles.between.species",random="yes",
   focus="species", species="Sap")
 # for both focal node and species
   retrieve.angles(evo.p,wishlist="angles.between.species",random="yes",
   focus="both", node=68, species="Sap")
 # without any specific requirement
   retrieve.angles(evo.p,wishlist="angles.between.species",random="yes",
   focus="none")
## End(Not run)
```

50 RRphylo

RRphylo

Evolutionary rates computation along phylogenies

## **Description**

The function RRphylo (Castiglione et al. 2018) performs the phylogenetic ridge regression. It takes a tree and a vector of tip data (phenotypes) as entries, calculates the regularization factor, produces the matrices of tip to root (makeL), and node to root distances (makeL1), the vector of ancestral state estimates, the vector of predicted phenotypes, and the rates vector for all the branches of the tree. For multivariate data, rates are given as both one vector per variable, and as a multivariate vector obtained by computing the Euclidean Norm of individual rate vectors.

#### Usage

RRphylo(tree,y,cov=NULL,rootV=NULL,aces=NULL,x1=NULL,aces.x1=NULL,clus=0.5)

#### **Arguments**

tree	a phylogenetic tree. The tree needs not to be ultrametric or fully dichotomous.
у	either a single vector variable or a multivariate dataset. In any case, y must be named. In case of categorical variable, this should be supplied to the function as a numeric vector.
cov	the covariate to be indicated if its effect on the rates must be accounted for. In this case residuals of the covariate versus the rates are used as rates. 'cov' must be as long as the number of nodes plus the number of tips of the tree, which can be obtained by running RRphylo on the covariate as well, and taking the vector of ancestral states and tip values to form the covariate, as in the example below. See RRphylo vignette - covariate for details.
rootV	phenotypic value (values if multivariate) at the tree root. If rootV=NULL the function 'learns' about the root value from the 10% tips being closest in time to the tree root, weighted by their temporal distance from the root itself (close tips phenotypes weigh more than more distant tips).
aces	a named vector (or matrix if y is multivariate) of ancestral character values at nodes. Names correspond to the nodes in the tree. See RRphylo vignette - aces for details.
x1	the additional predictor(s) to be indicated to perform the multiple version of RRphylo. 'x1' vector/matrix must be as long as the number of nodes plus the number of tips of the tree, which can be obtained by running RRphylo on the predictors (separately for each predictor) as well, and taking the vector of ancestral states and tip values to form the x1. See RRphylo vignette - predictor for details.
aces.x1	a named vector/matrix of ancestral character values at nodes for $x1$ . It must be indicated if both aces and $x1$ are specified. Names/rownames correspond to the nodes in the tree.
clus	the proportion of clusters to be used in parallel computing (only if y is multivariate). To run the single-threaded version of RRphylo set $clus = 0$ .

RRphylo 51

#### Value

**tree** the tree used by RRphylo. The fully dichotomous version of the tree argument. For trees with polytomies, the tree is resolved by using multi2di function in the package **ape**. Note, tip labels are ordered according to their position in the tree.

**tip.path** a n\*m matrix, where n=number of tips and m=number of branches (i.e. 2\*n-1). Each row represents the branch lengths along a root-to-tip path.

**node.path** a n \* n matrix, where n=number of internal branches. Each row represents the branch lengths along a root-to-node path.

**rates** single rate values computed for each branch of the tree. If y is a single vector variable, rates are equal to multiple.rates. If y is a multivariate dataset, rates are computed as the square root of the sum of squares of each row of \$multiple.rates.

aces the phenotypes reconstructed at nodes.

**predicted.phenotypes** the vector of estimated tip values. It is a matrix in the case of multivariate data.

**multiple.rates** a n \* m matrix, where n= number of branches (i.e. n\*2-1) and m = number of variables. For each branch, the column entries represent the evolutionary rate.

**lambda** the regularization factor fitted within RRphylo by the inner function optL. With multivariate data, several optL runs are performed. Hence, the function provides a single lambda for each individual variable.

**ace.values** if aces are specified, the function returns a dataframe containing the corresponding node number on the RRphylo tree for each node, along with estimated values.

x1.rate if x1 is specified, the function returns the partial regression coefficient for x1.

#### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

#### References

Castiglione, S., Tesone, G., Piccolo, M., Melchionna, M., Mondanaro, A., Serio, C., Di Febbraro, M., & Raia, P.(2018). A new method for testing evolutionary rate variation and shifts in phenotypic evolution. *Methods in Ecology and Evolution*, 9: 974-983.doi:10.1111/2041-210X.12954

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52 RRphylo

on the tempo and mode of brain size evolution in mammals. *Biological Journal of the Linnean Society*,132: 221-231. doi:10.1093/biolinnean/blaa186

```
## Not run:
data("DataOrnithodirans")
DataOrnithodirans$treedino->treedino
DataOrnithodirans$massdino->massdino
# Case 1. "RRphylo" without accounting for the effect of a covariate
RRphylo(tree=treedino,y=massdino)->RRcova
# Case 2. "RRphylo" accounting for the effect of a covariate
# "RRphylo" on the covariate in order to retrieve ancestral state values
c(RRcova$aces,massdino)->cov.values
c(rownames(RRcova$aces), names(massdino))->names(cov.values)
RRphylo(tree=treedino,y=massdino,cov=cov.values)->RR
# Case 3. "RRphylo" specifying the ancestral states
data("DataCetaceans")
DataCetaceans$treecet->treecet
DataCetaceans$masscet->masscet
DataCetaceans$brainmasscet->brainmasscet
DataCetaceans$aceMyst->aceMyst
RRphylo(tree=treecet,y=masscet,aces=aceMyst)->RR
# Case 4. Multiple "RRphylo"
library(ape)
drop.tip(treecet, treecet$tip.label[-match(names(brainmasscet), treecet$tip.label)])->treecet.multi
masscet[match(treecet.multi$tip.label,names(masscet))]->masscet.multi
RRphylo(tree=treecet.multi, y=masscet.multi)->RRmass.multi
RRmass.multi$aces[,1]->acemass.multi
c(acemass.multi,masscet.multi)->x1.mass
RRphylo(tree=treecet.multi,y=brainmasscet,x1=x1.mass)->RR
# Case 5. Categorical and multiple "RRphylo" with 2 additional predictors
library(phytools)
set.seed(1458)
rtree(50)->tree
fastBM(tree)->y
jitter(y)*10->y1
rep(1,length(y))->y2
y2[sample(1:50,20)]<-2
names(y2)<-names(y)</pre>
c(RRphylo(tree,y1)$aces[,1],y1)->x1
```

RRphylo-defunct 53

RRphylo-defunct

Defunct functions in Package RRphylo

# Description

These functions are no longer available.

• swap.phylo: This function is defunct. Please check overfitRR, instead.

#### Usage

```
swap.phylo()
```

RRphylo-deprecated

Deprecated functions in Package RRphylo

# Description

These functions still work but will be removed (defunct) in the next version.

scaleTree

Phylogenetic tree calibration

# Description

The function is a wrapper around the functions "scalePhylo", "assign.ages", and "assign.brlen" written by Gene Hunt (http://paleobiology.si.edu/staff/individuals/hunt.cfm). It rescales tree branch lengths according to given calibration dates.

## Usage

```
scaleTree(tree, tip.ages=NULL, node.ages=NULL,min.branch=NULL)
```

54 scaleTree

## Arguments

a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

tip.ages

a named vector including the ages (i.e. distance from the youngest tip within the tree) of the tips to be changed. If unspecified, the function assumes all the tips are correctly placed with respect to the root.

node.ages

a named vector including the ages (i.e. distance from the youngest tip within the

tree) of the nodes to be changed. If no calibration date for nodes is supplied, the tree root is fixed and the function shifts node position only where needed to fit

tip ages.

min.branch has been deprecated.

#### Value

Rescaled phylogentic tree with tip labels ordered according to their position in the tree.

# Author(s)

Silvia Castiglione, Pasquale Raia, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

#### See Also

scaleTree vignette

```
library(ape)
library(phytools)
data("DataFelids")
DataFelids$treefel->tree
max(nodeHeights(tree))->H
#### Example 1 ####
rep(0,4)->tipAges
names(tipAges)<-tips(tree,146)</pre>
scaleTree(tree,tipAges)->treeS1
edge.col<-rep("black",nrow(tree$edge))</pre>
edge.col[which(treeS1$edge[,2]%in%getDescendants(treeS1,146))]<-"red"
layout(2:1)
plot(tree,edge.color = edge.col,show.tip.label=FALSE)
plot(treeS1,edge.color = edge.col,show.tip.label=FALSE)
#### Example 2 ####
nodeAges < -c(23.5, 15.6)
names(nodeAges) < -c(85, 139)
```

search.conv 55

```
scaleTree(tree, node.ages=nodeAges)->treeS2
edge.col<-rep("black",nrow(tree$edge))
edge.col[which(treeS1$edge[,2]%in%c(getDescendants(treeS1,85),
                                     getDescendants(treeS1,139)))]<-"red"</pre>
layout(2:1)
plot(tree,edge.color = edge.col,show.tip.label=FALSE)
nodelabels(bg="w",frame="n",node=c(85,139),col="green")
plot(treeS2,edge.color = edge.col,show.tip.label=FALSE)
nodelabels(bg="w",frame="n",node=c(85,139),col="green")
#### Example 3 ####
16->nodeAges
names(nodeAges)<-"145"
tipAges<-19
names(tipAges)<-tree$tip.label[1]</pre>
scaleTree(tree,tip.ages = tipAges,node.ages=nodeAges)->treeS3
edge.col<-rep("black",nrow(tree$edge))</pre>
edge.col[which(treeS3$edge[,2]%in%c(1,getMommy(tree,1),
                                     getDescendants(treeS3,145)))]<-"red"</pre>
layout(2:1)
plot(tree,edge.color = edge.col,show.tip.label=FALSE)
nodelabels(bg="w",frame="n",node=145,col="green")
plot(treeS3,edge.color = edge.col,show.tip.label=FALSE)
nodelabels(bg="w",frame="n",node=145,col="green")
```

search.conv

Searching for morphological convergence among species and clades

# Description

The function scans a phylogenetic tree looking for morphological convergence between entire clades or species evolving under specific states.

#### Usage

```
search.conv(RR=NULL,tree=NULL,y,nodes=NULL,state=NULL,aceV=NULL,
    min.dim=NULL,max.dim=NULL,min.dist=NULL,declust=FALSE,nsim=1000,rsim=1000,
    clus=.5,filename=NULL)
```

#### **Arguments**

RR an object produced by RRphylo. This is not indicated if convergence among

states is tested.

tree a phylogenetic tree. The tree needs not to be ultrametric or fully dichotomous.

This is not indicated if convergence among clades is tested.

56 search.conv

a multivariate phenotype. The object y should be either a matrix or dataframe У with species names as rownames. node pair to be tested. If unspecified, the function automatically searches for nodes convergence among clades. Notice the node number must refer to the dichotomic version of the original tree, as produced by RRphylo. state the named vector of tip states. The function tests for convergence within a single state or among different states (this latter case is especially meant to test for iterative evolution as for example the appearance of repeated morphotypes into different clades). In both cases, the state for non-focal species (i.e. not belonging to any convergent group) must be indicated as "nostate". phenotypic values at internal nodes. The object aceV should be either a matrix aceV or dataframe with nodes (referred to the dichotomic version of the original tree, as produced by RRphylo) as rownames. If aceV are not indicated, ancestral phenotypes are estimated via RRphylo. min.dim the minimum size of the clades to be compared. When nodes is indicated, it is the minimum size of the smallest clades in nodes, otherwise it is set at one tenth max.dim the maximum size of the clades to be compared. When nodes is indicated, it is min.dim\*2 if the largest clade in nodes is smaller than this value, otherwise it corresponds to the size of the largest clade. Without nodes it is set at one third of the tree size. min.dist the minimum distance between the clades to be compared. When nodes is indicated, it is the distance between the pair. Under the automatic mode, the user can choose whether time distance or node distance (i.e. the number of nodes intervening between the pair) should be used. If time distance has to be considered, min.dist should be a character argument containing the word "time" and then the actual time distance to be used. The same is true for node distance, but the word "node" must precede the node distance to be used. For example, if the user want to test only clades more distant than 10 time units, the argument should be "time10". If clades separated by more than 8 nodes has to be tested, the argument min. dist should be "node8". If left unspecified, it automatically searches for convergence between clades separated by a number of nodes bigger than one tenth of the tree size. declust if species under a given state (or a pair of states) to be tested for convergence are phylogenetically closer than expected by chance, trait similarity might depend on proximity rather than true convergence. In this case, by setting declust = TRUE, tips under the focal state (or states) are removed randomly until clustering disappears. A minimum of 3 species per state is enforced to remain anyway. nsim number of simulations to perform sampling within the theta random distribution. It is set at 1000 by default. number of simulations to be performed to produce the random distribution of rsim theta values. It is set at 1000 by default. clus the proportion of clusters to be used in parallel computing. To run the singlethreaded version of search. conv set clus = 0. filename is deprecated. search.conv does not return plots anymore, check the function

plotConv instead.

search.conv 57

#### Value

If convergence between clades is tested, the function returns a list including:

- **\$node pairs**: a dataframe containing for each pair of nodes:
  - ang.bydist.tip: the mean theta angle between clades divided by the time distance.
  - ang.conv: the mean theta angle between clades plus the angle between aces, divided by the time distance.
  - ang.ace: the angle between aces.
  - ang.tip: the mean theta angle between clades.
  - nod.dist: the distance intervening between clades in terms of number of nodes.
  - time.dist: the time distance intervening between the clades.
  - p.ang.bydist: the p-value computed for ang.bydist.tip.
  - p.ang.conv: the p-value computed for ang.conv.
  - clade.size: the size of clades.
- **\$node pairs comparison**: pairwise comparison between significantly convergent pairs (all pairs if no instance of significance was found) performed on the distance from group centroids (the mean phenotype per clade).
- **\$average distance from group centroids**: smaller average distances mean less variable phenotypes within the pair.

If convergence between (or within a single state) states is tested, the function returns a list including:

- state.res a dataframe including for each pair of states (or single state):
  - ang.state: the mean theta angle between species belonging to different states (or within a single state).
  - ang.state.time: the mean of theta angle between species belonging to different states (or within a single state) divided by time distance.
  - p.ang.state: the p-value computed for ang.state.
  - p.ang.state.time: the p-value computed for ang.state.time.
- plotData a dataframe including data to plot the results via plotConv

Author(s)

Silvia Castiglione, Carmela Serio, Pasquale Raia, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto, Paolo Piras, Davide Tamagnini

# References

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58 search.shift

## See Also

```
search.conv vignette
plotConv vignette
```

# Examples

```
## Not run:
data("DataFelids")
DataFelids$PCscoresfel->PCscoresfel
DataFelids$treefel->treefel
DataFelids$statefel->statefel
cc<- 2/parallel::detectCores()

RRphylo(treefel,PCscoresfel,clus=cc)->RRfel

## Case 1. searching convergence between clades
# by setting min.dist as node distance
search.conv(RR=RRfel, y=PCscoresfel, min.dim=5, min.dist="node9",clus=cc)
# by setting min.dist as time distance
search.conv(RR=RRfel, y=PCscoresfel, min.dim=5, min.dist="time38",clus=cc)

## Case 2. searching convergence within a single state
search.conv(tree=treefel, y=PCscoresfel, state=statefel,declust=TRUE,clus=cc)

## End(Not run)
```

search.shift

Locating shifts in phenotypic evolutionary rates

#### **Description**

The function search.shift (*Castiglione et al. 2018*) tests whether individual clades or isolated tips dispersed through the phylogeny evolve at different RRphylo rates as compared to the rest of the tree. Instances of rate shifts may be automatically found.

## Usage

```
search.shift(RR, status.type = c("clade", "sparse"),node = NULL, state
= NULL, cov = NULL, nrep = 1000, f = NULL,filename=NULL)
```

# Arguments

```
RR an object fitted by the function RRphylo.

status.type whether the "clade" or "sparse" condition must be tested.
```

search.shift 59

node under the "clade" condition, the node (clade) to be tested for the rate shift. When multiple nodes are tested, they need to be written as in the example below. If node is left unspecified, the function performs under the 'auto-recognize' feature, meaning it will automatically test individual clades for deviation of their rates from the background rate of the rest of the tree (see details). the state of the tips specified under the "sparse" condition. state the covariate vector to be indicated if its effect on rate values must be accounted COV for. Contrary to RRphylo, cov needs to be as long as the number of tips of the tree. the number of simulations to be performed for the rate shift test, by default nrep nrep is set at 1000. f the size of the smallest clade to be tested. By default, nodes subtending to one tenth of the tree tips are tested. filename is deprecated. search. shift does not return plots anymore, check the function plotShift instead.

#### **Details**

The function search. shift takes the object produced by RRphylo. Two different conditions of rate change can be investigated. Under the "clade" condition the vector of node or nodes subjected to the shift must be provided. Alternatively, under the "sparse" case the (named) vector of states (indicating which tips are or are not evolving under the rate shift according to the tested hypothesis) must be indicated. In the "clade" case, the function may perform an 'auto-recognize' feature. Under such specification, the function automatically tests individual clades (from clades as large as one half of the tree down to a specified clade size) for deviation of their rates from the background rate of the rest of the tree, which is identical to the "clade" case. An inclusive clade with significantly high rates is likely to include descending clades with similarly significantly high rates. Hence, with 'auto-recognize' the search, shift function is written as to scan clades individually and to select only the node subtending to the highest difference in mean absolute rates as compared to the rest of the tree. Caution must be put into interpreting the 'auto-recognize' results. For instance, a clade with small rates and another with large rates could be individuated even under BM. This does not mean these clades are actual instances for rate shifts. Clades must be tested on their own without the 'auto-recognize' feature, which serves as guidance to the investigator, when no strong a priori hypothesis to be tested is advanced. The 'auto-recognize' feature is not meant to provide a test for a specific hypothesis. It serves as an optional guidance to understand whether and which clades show significantly large or small rates as compared to the rest of the tree. Individual clades are tested at once, meaning that significant instances of rate variation elsewhere on the tree are ignored. Conversely, running the "clade" condition without including the 'auto-recognize' feature, multiple clades presumed to evolve under the same shift are tested together, meaning that their rates are collectively contrasted to the rest of the tree, albeit they can additionally be compared to each other upon request. Under both the "clade" and "sparse" conditions, multiple clades could be specified at once, and optionally tested individually (for deviation of rates) against the rates of the rest of the tree and against each other. Regardless of which condition is specified, the function output produces the real difference of means, and their significance value.

60 search.shift

#### Value

Under all circumstances, search. shift provides a vector of \$rates. If 'cov' values are provided, rates are regressed against the covariate and the residuals of such regression form the vector \$rates. Otherwise, \$rates are the same rates as with the RR argument.

Under "clade" case without specifying nodes (i.e. 'auto-recognize') a list including:

**\$all.clades** for each detected node, the data-frame includes the average rate difference (computed as the mean rate over all branches subtended by the node minus the average rate for the rest of the tree) and the probability that it do represent a real shift. Probabilities are contrasted to simulations shuffling the rates across the tree branches for a number of replicates specified by the argument nrep. Note that the p-values refer to the number of times the real average rates are larger (or smaller) than the rates averaged over the rest of the tree, divided by the number of simulations. Hence, large rates are significantly larger than the rest of the tree (at alpha = 0.05), when the probability is > 0.975; and small rates are significantly small for p < 0.025.

**\$single.clades** the same as with 'all.clades' but restricted to the largest/smallest rate values along a single clade (i.e. nested clades with smaller rate shifts are excluded). Large rates are significantly larger than the rest of the tree (at alpha = 0.05), when the probability is > 0.975; and small rates are significantly small for p < 0.025.

Under "clade" condition by specifying the node argument:

**\$all.clades.together** if more than one node is tested, this specifies the average rate difference and the significance of the rate shift, by considering all the specified nodes as evolving under a single rate. As with the 'auto-recognize' feature, large rates are significantly larger than the rest of the tree (at alpha = 0.05), when the probability is > 0.975; and small rates are significantly small for p < 0.025.

**\$single.clades** this gives the significance for individual clades, tested separately. As previously, large rates are significantly larger than the rest of the tree (at alpha = 0.05), when the probability is > 0.975; and small rates are significantly small for p < 0.025.

Under the "sparse" condition:

**\$state.results** for each state, the data-frame includes the average rate difference (computed as the mean rate over all leaves evolving under a given state, minus the average rate for each other state or the rest of the tree) and the probability that the shift is real. Large rates are significantly larger (at alpha = 0.05), when the probability is > 0.975; and small rates are significantly small for p < 0.025. States are compared pairwise.

#### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

#### References

Castiglione, S., Tesone, G., Piccolo, M., Melchionna, M., Mondanaro, A., Serio, C., Di Febbraro, M., & Raia, P.(2018). A new method for testing evolutionary rate variation and shifts in phenotypic evolution. *Methods in Ecology and Evolution*, 9: 974-983.doi:10.1111/2041-210X.12954

#### See Also

```
search.shift vignette plotShift vignette
```

# Examples

```
## Not run:
data("DataOrnithodirans")
DataOrnithodirans$treedino->treedino
DataOrnithodirans$massdino->massdino
DataOrnithodirans$statedino->statedino
RRphylo(tree=treedino,y=massdino)->dinoRates
# Case 1. Without accounting for the effect of a covariate
# Case 1.1 "clade" condition
# with auto-recognize
search.shift(RR=dinoRates,status.type="clade")
# testing two hypothetical clades
search.shift(RR=dinoRates, status.type="clade", node=c(696,746))
# Case 1.2 "sparse" condition
# testing the sparse condition.
search.shift(RR=dinoRates, status.type= "sparse", state=statedino)
# Case 2. Accounting for the effect of a covariate
# Case 2.1 "clade" condition
search.shift(RR=dinoRates, status.type= "clade", cov=massdino)
# Case 2.2 "sparse" condition
search.shift (RR=dinoRates, status.type="sparse", state=statedino, cov=massdino)\\
## End(Not run)
```

search.trend

Searching for evolutionary trends in phenotypes and rates

# Description

This function searches for evolutionary trends in the phenotypic mean and the evolutionary rates for the entire tree and individual clades.

#### Usage

```
search.trend(RR,y,x1=NULL,x1.residuals = FALSE,
node=NULL,cov=NULL,nsim=100,clus=0.5,ConfInt=NULL,filename=NULL)
```

#### **Arguments**

RR an object produced by RRphylo.

y the named vector (or matrix if multivariate) of phenotypes.

x1 the additional predictor to be specified if the RR object has been created using

an additional predictor (i.e. multiple version of RRphylo). 'x1' vector must be as long as the number of nodes plus the number of tips of the tree, which can be obtained by running RRphylo on the predictor as well, and taking the vector of ancestral states and tip values to form the x1. Note: only one predictor at once

can be specified.

x1.residuals logical specifying whether the residuals of regression between y and x1 should

be inspected for a phenotypic trend (see details and examples below). Default is

FALSE.

node the node number of individual clades to be specifically tested and contrasted to

each other. It is NULL by default. Notice the node number must refer to the

dichotomic version of the original tree, as produced by RRphylo.

cov the covariate values to be specified if the RR object has been created using a

covariate for rates calculation. As for RRphylo, 'cov' must be as long as the number of nodes plus the number of tips of the tree, which can be obtained by running RRphylo on the covariate as well, and taking the vector of ancestral

states and tip values to form the covariate (see the example below).

nsim number of simulations to be performed. It is set at 100 by default.

clus the proportion of clusters to be used in parallel computing. To run the single-

threaded version of search. trend set clus = 0.

ConfInt is deprecated.

filename is deprecated. search. trend does not return plots anymore, check the function

plotTrend instead.

#### Details

The function simultaneously returns the regression of phenotypes and phenotypic evolutionary rates against age tested against Brownian motion simulations to assess significance. To this aim rates are rescaled in the 0-1 range and then logged. To assess significance, slopes are compared to a family of simulated slopes (BMslopes, where the number of simulations is equal to nsim), generated under the Brownian motion, using the fastBM function in the package **phytools**. Individual nodes are compared to the rest of the tree in different ways depending on whether phenotypes or rates (always unscaled in this case) versus age regressions are tested. With the former, the regression slopes for individual clades and the slope difference between clades is contrasted to slopes obtained through Brownian motion simulations. For the latter, regression models are tested and contrasted to each other referring to estimated marginal means, by using the emmeans function in the package **emmeans**.

The multiple regression version of RRphylo allows to incorporate the effect of an additional predictor in the computation of evolutionary rates without altering the ancestral character estimation. Thus, when a multiple RRphylo output is fed to search. trend, the predictor effect is accounted for on the absolute evolutionary rates, but not on the phenotype. However, in some situations the user might want to factor out the predictor effect on phenotypes as well. Under the latter circumstance,

by setting the argument x1.residuals = TRUE, the residuals of the response to predictor regression are used as to represent the phenotype.

#### Value

The function returns a list object containing:

**\$trends.data** a 'RRphyloList' object including:

- 1. \$phenotypeVStime: a data frame of phenotypic values (or y versus x1 regression residuals if x1.residuals=TRUE) and their distance from the tree root for each node (i.e. ancestral states) and tip of the tree.
- 2. \$absrateVStime: a data frame of RRphylo rates and the distance from the tree root (age). If y is multivariate, it also includes the multiple rates for each y vector. If node is specified, each branch is classified as belonging to an indicated clade.
- 3. \$rescaledrateVStime: a data frame of rescaled RRphylo rates and the distance from the tree root (age). If y is multivariate, it also includes the multiple rates for each y vector. If node is specified, each branch is classified as belonging to an indicated clade. NAs correspond either to very small values or to outliers which are excluded from the analysis.

**\$phenotypic.regression** results of phenotype (y versus x1 regression residuals) versus age regression. It reports a p-value for the regression slope between the variables (p.real), a p-value computed contrasting the real slope to Brownian motion simulations (p.random), and a parameter indicating the deviation of the phenotypic mean from the root value in terms of the number of standard deviations of the trait distribution (dev). dev is 0 under Brownian Motion. Only p.random should be inspected to assess significance.

**\$rate.regression** results of the rates (rescaled absolute values) versus age regression. It reports a p-value for the regression between the variables (p.real), a p-value computed contrasting the real slope to Brownian motion simulations (p.random), and a parameter indicating the ratio between the range of phenotypic values and the range of such values halfway along the tree height, divided to the same figure under Brownian motion (spread). spread is 1 under Brownian Motion. Only p.random should be inspected to assess significance.

**\$ConfInts** a 'RRphyloList' object including the 95% confidence intervals around regression slopes of phenotypes and rates (both rescaled and unscaled absolute rates) produced according to the Brownian motion model of evolution.

If specified, individual nodes are tested as the whole tree, the results are summarized in the objects:

**\$node.phenotypic.regression** results of phenotype (or y versus x1 regression residuals) versus age regression through node. It reports the slope for the regression between the variables at node (slope), a p-value computed contrasting the real slope to Brownian motion simulations (p.random), the difference between estimated marginal means predictions for the group and for the rest of the tree (emm.difference), and a p-value for the emm.difference (p.emm).

**\$node.rate.regression** results of the rates (absolute values) versus age regression through node. It reports the difference between estimated marginal means predictions for the group and for the rest of the tree (emm.difference), a p-value for the emm.difference (p.emm), the regression slopes for the group (slope.node) and for the rest of the tree (slope.others), and a p-value for the difference between such slopes (p.slope).

If more than one node is specified, the object **\$group.comparison** reports the same results as \$node.phenotypic.regression and \$node.rate.regression obtained by comparing individual clades to each other.

#### Author(s)

Silvia Castiglione, Carmela Serio, Pasquale Raia, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

#### References

Castiglione, S., Serio, C., Mondanaro, A., Di Febbraro, M., Profico, A., Girardi, G., & Raia, P. (2019) Simultaneous detection of macroevolutionary patterns in phenotypic means and rate of change with and within phylogenetic trees including extinct species. *PLoS ONE*, 14: e0210101. https://doi.org/10.1371/journal.pone.0210101

#### See Also

```
search.trend vignette
plotTrend vignette
plotTrend
```

```
## Not run:
data("DataOrnithodirans")
DataOrnithodirans$treedino->treedino
DataOrnithodirans$massdino->massdino
cc<- 2/parallel::detectCores()</pre>
# Extract Pterosaurs tree and data
library(ape)
extract.clade(treedino,746)->treeptero
massdino[match(treeptero$tip.label,names(massdino))]->massptero
massptero[match(treeptero$tip.label,names(massptero))]->massptero
# Case 1. "RRphylo" whitout accounting for the effect of a covariate
RRphylo(tree=treeptero,y=log(massptero))->RRptero
# Case 1.1. "search.trend" whitout indicating nodes to be tested for trends
search.trend(RR=RRptero, y=log(massptero), nsim=100, clus=cc,cov=NULL,node=NULL)
# Case 1.2. "search.trend" indicating nodes to be specifically tested for trends
search.trend(RR=RRptero, y=log(massptero), nsim=100, node=143, clus=cc,cov=NULL)
# Case 2. "RRphylo" accounting for the effect of a covariate
# "RRphylo" on the covariate in order to retrieve ancestral state values
RRphylo(tree=treeptero,y=log(massptero))->RRptero
c(RRptero$aces,log(massptero))->cov.values
names(cov.values)<-c(rownames(RRptero$aces), names(massptero))</pre>
RRphylo(tree=treeptero,y=log(massptero),cov=cov.values)->RRpteroCov
# Case 2.1. "search.trend" whitout indicating nodes to be tested for trends
search.trend(RR=RRpteroCov, y=log(massptero), nsim=100, clus=cc,cov=cov.values)
```

setBM 65

```
# Case 2.2. "search.trend" indicating nodes to be specifically tested for trends
search.trend(RR=RRpteroCov, y=log(massptero), nsim=100, node=143, clus=cc,cov=cov.values)
# Case 3. "search.trend" on multiple "RRphylo"
data("DataCetaceans")
DataCetaceans$treecet->treecet
DataCetaceans$masscet->masscet
DataCetaceans$brainmasscet->brainmasscet
DataCetaceans$aceMyst->aceMyst
drop.tip(treecet, treecet$tip.label[-match(names(brainmasscet), treecet$tip.label)])->treecet.multi
masscet[match(treecet.multi$tip.label,names(masscet))]->masscet.multi
RRphylo(tree=treecet.multi,y=masscet.multi)->RRmass.multi
RRmass.multi$aces[,1]->acemass.multi
c(acemass.multi,masscet.multi)->x1.mass
RRphylo(tree=treecet.multi,y=brainmasscet,x1=x1.mass)->RRmulti
# incorporating the effect of body size at inspecting trends in absolute evolutionary rates
search.trend(RR=RRmulti, y=brainmasscet,x1=x1.mass,clus=cc)
# incorporating the effect of body size at inspecting trends in both absolute evolutionary
# rates and phenotypic values (by using brain versus body mass regression residuals)
search.trend(RR=RRmulti, y=brainmasscet,x1=x1.mass,x1.residuals=TRUE,clus=cc)
## End(Not run)
```

setBM

Producing simulated phenotypes with trends

#### **Description**

The function setBM is wrapper around **phytools** fastBM function, which generates BM simulated phenotypes with or without a trend.

# Usage

```
setBM(tree, nY = 1, s2 = 1, a = 0, type = c("", "brown","trend",
   "drift"), trend.type = c("linear", "stepwise"),tr = 10, t.shift = 0.5,
   es=2, ds=1)
```

#### **Arguments**

tree	a phylogenetic tree.
nY	the number of phenotypes to simulate.
s2	value of the Brownian rate to use in the simulations.
а	the phenotype at the tree root.

66 setBM

type	the type of phenotype to simulate. With the option "brown" the phenotype will have no trend in the phenotypic mean or in the rate of evolution. A variation in the phenotypic mean over time (a phenotypic trend) is obtained by selecting the option "drift". A trend in the rate of evolution produces an increased variance in the residuals over time. This is obtained by specifying the option "trend".
trend.type	two kinds of heteroscedastic residuals are generated under the "trend" type. The option "linear" produces an exponential linear increase (or decrease) in heteroscedasticity, whereas the "stepwise" option produces an increase (or decrease) after a specified point in time.
tr	the intensity of the trend with the "stepwise" option is controlled by the tr argument. The scalar tr is the multiplier of the branches extending after the shift point as indicated by t.shift.
t.shift	the relative time distance from the tree root where the stepwise change in the rate of evolution is indicated to apply.
es	when trend.type="linear", es is a scalar representing the exponent at which the evolutionary time (i.e. distance from the root) scales to change to phenotypic variance. With $es=1$ the phenotypic rate will be trendless, with $es<1$ the variance of the phenotypes will decrease exponentially towards the present and the other way around with $es>1$ .
ds	a scalar indicating the change in phenotypic mean in the unit time, in type="drift" case. With ds = 0 the phenotype will be trendless, with ds < 0 the phenotypic mean will decrease exponentially towards the present and the other way around with ds > 0.

#### **Details**

Note that setBM differs from fastBM in that the produced phenotypes are checked for the existence of a temporal trend in the phenotype. The user may specify whether she wants trendless data (option "brown"), phenotypes trending in time (option "drift"), or phenotypes whose variance increases/decreases exponentially over time, consistently with the existence of a trend in the rate of evolution (option "trend"). In the latter case, the user may indicate the intensity of the trend (by applying different values of es), and whether it should occur after a given proportion of the tree height (hence a given point back in time, specified by the argument t.shift). Trees in setBM are treated as non ultrametric. If an ultrametric tree is fed to the function, setBM alters slightly the leaf lengths multiplying randomly half of the leaves by 1 \* 10e-3, in order to make it non-ultrametric.

#### Value

Either an object of class 'array' containing a single phenotype or an object of class 'matrix' of n phenotypes as columns, where n is indicated as nY = n.

#### Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

sig2BM 67

## **Examples**

```
data("DataOrnithodirans")
DataOrnithodirans$treedino->treedino

setBM(tree=treedino, nY= 1, type="brown")
setBM(tree=treedino, nY= 1, type="drift", ds=2)
setBM(tree=treedino, nY= 1, type="trend", trend.type="linear", es=2)
```

sig2BM

Brownian Motion rate computation

#### **Description**

The function computes rate of phenotypic evolution along a phylogeny assuming Brownian Motion model of evolution.

#### Usage

```
sig2BM(tree,y)
```

# Arguments

tree a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

y either a single vector variable or a multivariate dataset. In any case, y must be named.

#### Value

The Brownian Motion rate of phenotypic evolution for each variable in y.

## Author(s)

Pasquale Raia, Silvia Castiglione

```
### Univariate data ###
data(DataCetaceans)
DataCetaceans$treecet->treecet
DataCetaceans$masscet->masscet
sig2BM(tree=treecet, y=masscet)

### Multivariate data ###
data(DataUng)
DataUng$treeung->treeung
DataUng$PCscoresung->PCscores
sig2BM(tree=treeung, y=PCscores)
```

68 sizedsubtree

sizedsubtree	Find a node subtending to a clade of desired size	

# **Description**

The function sizedsubtree scans a phylogentic tree to randomly find nodes subtending to a subtree of desired minimum size, up to one half of the tree size (number of tips).

# Usage

```
sizedsubtree(tree,Size=NULL,time.limit=10)
```

# **Arguments**

tree a phylogenetic tree.

Size the desired size of the tree subtending to the extracted node. By default, the

minimum tree size is set at one tenth of the tree size (i.e. number of tips).

time.limit specifies a limit to the searching time, a warning message is thrown if the limit

is reached.

# **Details**

The argument time.limit sets the searching time. The algorithm stops if that limit is reached, avoiding recursive search when no solution is in fact possible.

#### Value

A node subtending to a subtree of desired minimum size.

## Author(s)

Pasquale Raia, Silvia Castiglione, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

```
data("DataOrnithodirans")
DataOrnithodirans$treedino->treedino
sizedsubtree(tree=treedino,Size=40)
```

StableTraitsR 69

StableTraitsR Run StableTraits from within R
--

# **Description**

This function runs StableTraits and StableTraitsSum (*Elliot and Mooers 2014*) from within the R environment and returns its output into the workspace.

#### Usage

StableTraitsR(tree,y,path,output=NULL,aces=NULL,argST=NULL)

#### **Arguments**

tree	a phylogenetic tree. The tree needs not to be either ultrametric or fully dichotomous.
У	a named vector of phenotypic trait.
path	the folder path where the StableTraits output will be stored. Notice that the input tree and data (modified automatically if the original tree is not fully dichotomous or if aces are specified) will be stored in this folder as well.
output	name of the output to be returned, if unspecified it will be named "output".
aces	a named vector of ancestral character values at nodes specified in advance. Names correspond to the nodes in the tree.
argST	a list of further arguments passed to StableTraits. If the argument has no value (for example "brownian") it must be specified as TRUE.
argSTS	list of further arguments passed to StableTraitsSum. If the argument has no value (for example "brownian") it must be specified as TRUE.

# Details

The StableTraits software is available at https://mickelliot.com/, along with instructions for compilation. Once it is installed, the user must set as R working directory the folder where the StableTraits software are installed. Further information about the arguments and outputs of StableTraits and StableTraitsSum can be found at https://mickelliot.com/. StableTraitsR automatically recognizes which Operating System is running on the computer (it has been tested successfully on MacOS and Windows machines).

#### Value

The function returns a 'list' containing the output of StableTraits and StableTraitsSum.

**\$progress** a table reporting the DIC and PRSF diagnostics.

**\$rates\_tree** a copy of the original tree with branch lengths set to the evolutionary rate imputed by the stable reconstruction. Specifically, each branch length is equal to the absolute difference in the stable reconstruction occurring on that branch divided by the square root of the input branch length.

70 swapONE

**\$rates** the original branch lengths, evolutionary rates, node height and (optionally) scaled branch lengths.

**\$aces** the median estimates of ancestral states and stable parameters along with the 95% credible interval.

**\$brownian\_tree** if "brownian" is TRUE in argSTS, a copy of the original tree with branch lengths set such that the Brownian motion reconstruction of the character on this tree is approximately the same as the stable ancestral reconstruction.

**\$ace.prior.values** if aces is specified, the function returns a dataframe containing the corresponding node number on the RRphylo tree for each node, the original (preset) and the estimated values, and the 95% credible interval.

#### Author(s)

Silvia Castiglione, Carmela Serio, Pasquale Raia

#### References

Elliot, M. G., & Mooers, A. Ø. (2014). Inferring ancestral states without assuming neutrality or gradualism using a stable model of continuous character evolution. *BMC evolutionary biology*, 14: 226. doi.org/10.1186/s12862-014-0226-8

#### **Examples**

```
## Not run:
library(ape)
library(phytools)

# Set as working directory the folder where StableTraits software are stored
# setwd("~/StableTraits")

dir.create("Analyses")
rtree(100)->tree
fastBM(tree)->y
c(1,2,3)->acev
sample(Ntip(tree)+seq(1:Nnode(tree)),3)->names(acev)
StableTraitsR(tree,y,path="Analyses/",output="my_output",aces=acev,
argST=list(iterations=500000,chains=4),argSTS=list(brownian=TRUE))->ST

## End(Not run)
```

swap0NE

Create alternative phylogenies from a given tree

#### **Description**

The function produces an alternative phylogeny with altered topology and branch length, and computes the Kuhner-Felsenstein (Kuhner & Felsenstein 1994) distance between original and 'swapped' tree.

swapONE 71

# Usage

```
swapONE(tree, node=NULL, si=0.5, si2=0.5, plot.swap=FALSE)
```

#### **Arguments**

tree	a phylogenetic tree. The tree needs not to be ultrametric or fully dichotomous.
node	if specified, the clades subtended by such node(s) are imposed to be monophyletic. In this case, the function can still swap tips <i>within</i> the clade.
si	the proportion of tips whose topologic arrangement will be swapped.
si2	the proportion of nodes whose age will be changed.
plot.swap	if TRUE, the function plots the swapped tree. Swapped positions appear in red. Nodes with altered ages appear in green.

#### **Details**

swapONE changes the tree topology and branch lengths. Up to half of the tips, and half of the branch lengths can be changed randomly. Each randomly selected node is allowed to move up to 2 nodes apart from its original position.

#### Value

The function returns a list containing the 'swapped' version of the original tree, and the Kuhner-Felsenstein distance between the trees. Note, tip labels are ordered according to their position in the tree.

#### Author(s)

Silvia Castiglione, Pasquale Raia, Carmela Serio, Alessandro Mondanaro, Marina Melchionna, Mirko Di Febbraro, Antonio Profico, Francesco Carotenuto

# References

Kuhner, M. K. & Felsenstein, J. (1994). A simulation comparison of phylogeny algorithms under equal and unequal evolutionary rates, *Molecular Biology and Evolution*, 11: 459-468.

72 tree.merger

tips Get descending tips

#### **Description**

The function returns the numbers or labels of tips descending from a given node.

#### Usage

```
tips(tree,node,labels=TRUE)
```

# Arguments

tree a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

node the number of focal node

labels if TRUE (default) the function returns the labels of descending tips.

#### Value

The tips, either labels or numbers depending on the argument labels, descending from the node.

#### Author(s)

Silvia Castiglione, Pasquale Raia, Carmela Serio

# **Examples**

data(DataOrnithodirans)
DataOrnithodirans\$treedino->treedino
tips(tree=treedino,node=677,labels=FALSE)
tips(tree=treedino,node=677,labels=TRUE)

tree.merger

Fast addition of tips and clades on an existing tree

# Description

The function attaches new tips and/or clades derived from a source phylogeny to a pre-existing backbone tree.

# Usage

```
tree.merger(backbone,data,source.tree=NULL,age.offset=NULL,tip.ages =
    NULL, node.ages = NULL,min.branch=NULL,plot=TRUE,filename=NULL)
```

tree.merger 73

#### **Arguments**

backbone the backbone tree to attach tips/clades on.

data a dataset including as columns:

- 1. bind = the tips/clades to be attached;
- 2. reference = the reference tip or clade where 'bind' must be attached;
- 3. poly = logical specifying if 'bind' and 'reference' should form a polytomous clade.

See details for further explanations.

source.tree the tree where 'bind' clades are to be extracted from. If no clade has to be

attached, it can be left unspecified.

age.offset if the most recent age (i.e. the maximum distance from the tree root) differs

between the source and the backbone trees, the "age.offset" is the difference between them in this exact order (source minus backbone). It is positive when the backbone tree attains younger age than the source tree, and vice-versa.

tip.ages as in scaleTree, a named vector including the ages (i.e. the time distance from

the youngest tip within the tree) of the tips. If unspecified, the function assumes all the tips on the backbone tree are correctly placed and places all the new tips at the maximum distance from the tree root (i.e. the present if the tips are extant).

node.ages as in scaleTree, a named vector including the ages (i.e. the time distance from

the youngest tip within the tree) of the nodes. The nodes must be defined by collating the names of the two phylogenetically furthest tips it subtends to, separated by a "-" (see examples). If no calibration date for nodes is supplied, the function may shift the node position back in time as to place new tips/clades and

to fit tip ages.

min.branch has been deprecated.

plot if TRUE, the function produces an interactive plotting device to check the placing

of each bind.

filename if plot=TRUE and provided a filename (with or without the path), the function

stores a pdf file showing the plot of the entire phylogeny.

#### **Details**

The function attaches tips and/or clades from the source tree to the backbone tree according to the data object. Within the latter, a clade, either to be bound or to be the reference, must be indicated by collating the names of the two phylogenetically furthest tips belonging to it, separated by a "-". Duplicated 'bind' produce error. Tips/clades set to be attached to the same 'reference' are considered to represent a polytomy. Tips set as 'bind' which are already on the backbone tree are removed from the latter and placed according to the 'reference'. See examples and vignette for clarifications.

#### Value

Merged phylogenetic tree.

74 tree.merger

#### Author(s)

Silvia Castiglione, Carmela Serio, Pasquale Raia

#### References

Castiglione, S., Serio, C., Mondanaro, A., Melchionna, M., & Raia, P. (2022). Fast production of large, time-calibrated, informal supertrees with tree.merger. *Palaeontology*, 65: e12588.https://doi.org/10.1111/pala.12588

#### See Also

tree.merger vignette; scaleTree vignette;

```
## Not run:
require(ape)
DataCetaceans$treecet->tree
{\tt data.frame(bind=c("Balaena\_mysticetus-Caperea\_marginata",
                   "Aetiocetus_weltoni",
                   "Saghacetus_osiris",
                   "Zygorhiza_kochii",
                   "Ambulocetus_natans",
                   "Kentriodon_pernix",
                   "Kentriodon_schneideri",
                   "Kentriodon_obscurus",
                   {\tt "Tursiops\_truncatus-Delphinus\_delphis"},
                   "Kogia_sima",
                   "Grampus_griseus"),
           reference=c("Fucaia_buelli-Aetiocetus_weltoni",
                        "Aetiocetus_cotylalveus",
                        "Fucaia_buelli-Tursiops_truncatus",
                        "Saghacetus_osiris-Fucaia_buelli",
                        "Dalanistes_ahmedi-Fucaia_buelli",
                        "Kentriodon_schneideri",
                        "Phocoena_phocoena-Delphinus_delphis",
                        "Kentriodon_schneideri",
                        "Stenella_attenuata-Stenella_longirostris",
                        "Kogia_breviceps",
                        "Globicephala_melas-Pseudorca_crassidens"),
           poly=c(FALSE,
                  FALSE,
                  FALSE,
                  FALSE,
                  FALSE,
                  FALSE,
                  FALSE,
                  FALSE,
                  FALSE,
                  FALSE,
                  FALSE))->dato
c(Aetiocetus_weltoni=28.0,
```

treeCompare 75

```
Saghacetus_osiris=33.9,
  Zygorhiza_kochii=34.0,
  Ambulocetus_natans=40.4,
  Kentriodon_pernix=15.9,
  Kentriodon_schneideri=11.61,
  Kentriodon_obscurus=13.65)->tip.ages
c("Ambulocetus_natans-Fucaia_buelli"=52.6,
   "Balaena_mysticetus-Caperea_marginata"=21.5)->node.ages
# remove some tips from the original tree and create a source tree
drop.tip(tree,c(names(tip.ages),
                 tips(tree,131)[-which(tips(tree,131)%in%
                               c("Caperea_marginata", "Eubalaena_australis"))],
                 tips(tree,195)[-which(tips(tree,195)=="Tursiops_aduncus")]))->backtree
drop.tip(tree,which(!tree$tip.label%in%c(names(tip.ages),
                                          tips(tree, 131),
                                          tips(tree,195))))->sourcetree
plot(backtree,cex=.6)
plot(sourcetree,cex=.6)
tree.merger(backbone=backtree,data=dato,source.tree=sourcetree,
            tip.ages=tip.ages,node.ages = node.ages, plot=TRUE)->treeM
## End(Not run)
```

treeCompare

Visualize the difference between phylogenetic trees

#### **Description**

The function scans a pair of phylogenetic trees to find topological differences.

# Usage

```
treeCompare(tree, tree1, plot = TRUE)
```

## **Arguments**

tree, tree1 a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

Generic name and specific epithet must be separated by '\_'.

plot if TRUE, the function produces an interactive plotting device to check differences

in species placement at the genus level.

## Value

The function returns a data-frame indicating for each un-matching species its sister species/clades on both trees.

76 treedataMatch

#### Author(s)

Silvia Castiglione, Carmela Serio, Antonella Esposito

## **Examples**

```
## Not run:
DataSimians$tree->tree
set.seed(22)
swapONE(tree,si=0.5)[[1]]->tree1
treeCompare(tree,tree1)
## End(Not run)
```

treedataMatch

Cross-reference tree and data

## Description

The function matches data names with tree tips. If either there is no data for a tip or it is not present on the tree, the function removes the entry from both.

#### Usage

```
treedataMatch(tree,y)
```

# **Arguments**

tree a phylogenetic tree. The tree needs not to be ultrametric and fully dichotomous.

y either a single vector variable or a multivariate dataset. In any case, y must be named.

#### Value

The function returns a list object. If no mismatch between tree and y is detected, the list only includes the matrix of y ordered according to the order of tips on the tree (\$y). If some tips on the tree are missing from y, they are removed from the phylogeny. Thus, the list also includes the pruned tree (\$tree) and the vector of dropped tips (\$removed.from.tree). Similarly, if some entries in y are missing from the tree, the list also includes the vector of mismatching entry names (\$removed.from.y). In this latter case, the first element of the list (\$y) does not include the entries \$removed.from.y, so that it perfectly matches the phylogeny.

# Author(s)

Silvia Castiglione, Pasquale Raia, Carmela Serio

treedataMatch 77

# Examples

data(DataCetaceans)
DataCetaceans\$treecet->treecet
DataCetaceans\$masscet->masscet
DataCetaceans\$brainmasscet->brainmasscet

treedataMatch(tree=treecet,y=masscet)
treedataMatch(tree=treecet,y=brainmasscet)

# **Index**

* RRphylo DataApes, 9 DataCetaceans, 10 DataFelids, 11 DataOrnithodirans, 11 DataSimians, 12 DataUng, 13	phyloclust, 33 phylolm, 32 plotConv, 34, 56, 57 plotRates, 36 plotShift, 38, 59 plotTrend, 39, 62, 64 polar.plot, 35
angle.matrix, 3	procSym, 7, 12, 44
conv.map, 6 cutPhylo, 8	random.evolvability.test,41 rate.map,43 relWarps,7,44
DataApes, 9 DataCetaceans, 10 DataFelids, 11 DataOrnithodirans, 11 DataSimians, 12 DataUng, 13 distNodes, 13	rescaleRR, 45 retrieve.angles, 46 RRphylo, 3, 4, 15, 27, 31, 34, 36–38, 40, 41, 43, 45, 47, 50, 50, 55, 58, 59, 62 RRphylo-defunct, 53 RRphylo-deprecated, 53 RRphylo-package, 3
evo.dir, 14, 46, 47	scaleTree, 53, 73 search.conv, 26, 34, 55
fix.poly, 17	search.shift, 26, 38, 58 search.trend, 26, 39, 61
<pre>getGenus, 19 getMommy, 20 getSis, 21</pre>	setBM, 65 sig2BM, 67 sizedsubtree, 68 StableTraitsR, 69
lollipoPlot, 21, 37	swap.phylo (RRphylo-defunct), 53 swapONE, 27, 28, 70
makeFossil, 22	
makeL, 23, 50	tips, 72
makeL1, 24, 50 MeanMatrixStatistics, 41	tree.merger, 72 treeCompare, 75 treedataMatch, 76
namesCompare, 25	
node.paths, 26	vcgBallPivoting, $6,43$
overfitRR, 26, 53	
PGLS_fossil, 26, 31	